

REMARKS/ARGUMENTS

Upon entry of the present amendment, claims 1-5, 17-19, 21-22, 24, 26, 29, 39, 41, 53, 55, 64, 88-90, and 92-100 are pending in the application, with claims 1-5, 17-19, 21-22, 24, 26, 29, 39, 41, 64, and 94-98 having been withdrawn from consideration for being directed to non-elected subject matter.

Claims 53, 55, 88-90, and 92 are amended, claims 54 and 91 are canceled, and new claims 99 and 100 are added in the present amendment. Support for the amendment to claim 53 can be found in the specification at, *e.g.*, paragraphs 0158 of the published application (US 2006/0258656 A1) and in original claim 55. Support for the amendment to claim 88 can be found in the specification at, *e.g.*, paragraphs 0171-0176 of the published application. Support for the amendment to claim 89 can be found in the specification at, *e.g.*, paragraph 0158 of the published application. Support for the amendment to claim 92 can be found in the specification at, *e.g.*, paragraphs 0160 and 0174 of the published application. Claim 55 is amended to reflect the amendment to claim 53 and the cancellation of claim 54. Claim 90 is amended to correct a grammatical error. Support for new claim 99 can be found in the specification at, *e.g.*, paragraphs 0160 and 0174 of the published application. Support for new claim 100 can be found in the specification at, *e.g.*, Example 21, including paragraph 0307 of the published application. No new matter is added by the present amendment.

Applicants address each of the Examiner's rejections and objections below in the order presented in the Office Action.

Claim Rejections under 35 U.S.C. 112, Second Paragraph

The Examiner has rejected claims 53-55 and 88-93 under 35 U.S.C. 112, second paragraph. The Examiner stated that the terms "anti-neoplastic agent," "hypoxic activator," and "alkylating agent" are indefinite. The Examiner suggested that this rejection could be overcome by amending the claims to recite a structurally defined hypoxic activator and specific anti-neoplastic agents. *See* p. 4, first paragraph of Office Action.

Without agreeing with the Examiner's position, applicants have amended independent claim 53 to recite the specific structurally-defined hypoxic activator formerly recited in dependent claim 55. Applicants have also amended independent claims 53 and 89 to indicate that the anti-neoplastic agent is an alkylating agent, and submit that, as discussed below, the term "alkylating agent" has a clear meaning and does not render the claims indefinite.

The Examiner stated that the term "alkylating agent" is unclear. *See* p. 4, 1st paragraph of the Office Action. The claims, as amended, recite the term "alkylating agent" in the context of anti-neoplastic agents, *i.e.*, the anti-neoplastic agent is an alkylating agent. Applicants submit that "alkylating agent," as used in the claims with reference to an anti-neoplastic agent, is a well known term of art that would be immediately recognized by a skilled artisan as conveying a specific meaning. *See, e.g., Cancer, Principles and Practice of Oncology*, 6th Edition, DeVita *et al.*, Lippencott Williams and Wilkins, Philadelphia, PA, pp. 363-376, 2001 (a copy of pages 363-376 are enclosed). DeVita *et al.* reports that antitumor alkylating agents react with DNA bases in cells to prevent cell replication. *See* p. 363. DeVita *et al.* also describes several classes of such alkylating agents, including agents such as cyclophosphamide, ifosfamide, melphalan, chlorambucil, and thiotepa (*see* pp. 363-366), which are also exemplified in the instant specification (*see* paragraph 0174 of the published application). The requirements of 35 U.S.C. 112, second paragraph, are satisfied if a person skilled in the field of the invention would reasonably understand the claim when read in the context of the specification. Here, the skilled person would understand what is meant by the term "alkylating agent," as used in the context of anti-neoplastic agents in both the specification and claims. Thus, use of the term "alkylating agent" does not render the claims indefinite.

The Examiner also stated that the breadth of the term "alkylating agent" prevents one from ascertaining the scope of the claims. *See* p. 4, first paragraph of the Office Action. However, breadth is not to be equated with indefiniteness. *See* MPEP §2173.04. Indefiniteness implies that one of skill would not be able to determine whether any particular anti-neoplastic agent is encompassed within the term "alkylating agent." As discussed above, the art and the specification make clear that an anti-neoplastic alkylating agent is an agent that reacts with DNA bases in cells to prevent cell replication. One of skill could readily determine that an alkylating

agent reacts with DNA bases in cells to prevent cell replication, and would be encompassed by the claims. Where the scope of the subject matter embraced by the claims is clear, then the claims comply with 35 U.S.C. 112, second paragraph. *See* MPEP §2173.04. Here, the scope of the claimed subject matter is clear because the term “alkylating agent,” as used to define the anti-neoplastic agents of the claimed invention, is a term of art whose meaning would be understood by the skilled artisan. Thus, the full scope of the claimed invention is clear and the claims are not indefinite.

Based on the arguments presented above, applicants respectfully request withdrawal of this ground of rejection.

Claim Rejection under 35 U.S.C. 112, First Paragraph

The Examiner has rejected claim 88 under 35 U.S.C. 112, first paragraph, because the specification allegedly fails to enable the full scope of the claim. The Examiner suggested that this rejection could be overcome by amending the claim to recite specific cancers identified in the specification. *See* p. 9, second full paragraph of the Office Action.

Without agreeing with the Examiner’s position, applicants have amended claim 88 to recite specific cancers identified in the application. Based on applicants’ disclosure, the skilled artisan could practice the full scope of the claimed invention without undue experimentation.

Based on the arguments presented above, applicants respectfully request withdrawal of this ground of rejection.

Claim Objections

The Examiner has objected to claims 53-55, and 88-93 as encompassing non-elected subject matter. The Examiner suggested that applicants amend the claims to the scope of elected subject matter set forth at pages 2-3 of the Office Action. *See* p. 10, first paragraph of the Office Action.

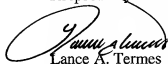
Without agreeing with the Examiner’s objections, applicants submit that the current claim amendments discussed above fully address the Examiner’s objections.

Applicants wish to point out that the Examiner's claim objections appear to include an objection to the recitation of various classes of anti-neoplastic agents, including "alkylating agents," previously set forth in claim 89. Applicants have not been able to identify any such restriction requirement in the Office Action of May 5, 2008, which set forth a restriction of the claims to the presently claimed hypoxic activator, and required election of a single compound for examination. *See* pp. 4-5 of the Office Action of May 5, 2008. Thus, applicants submit that the current claim amendments fully address the Examiner's objections to the inclusion of non-elected subject matter.

Based on the arguments presented above, applicants respectfully request withdrawal of the Examiner's claim objections.

If the Examiner believes a telephone conference would expedite prosecution of this application, please telephone the undersigned at 650-326-2400.

Respectfully submitted,



Lance A. Termes
Reg. No. 43,184

TOWNSEND and TOWNSEND and CREW LLP
Two Embarcadero Center, Eighth Floor
San Francisco, California 94111-3834
Tel: 650-326-2400
Fax: 415-576-0300
LAT:y1m
Attachment
61713992 v1

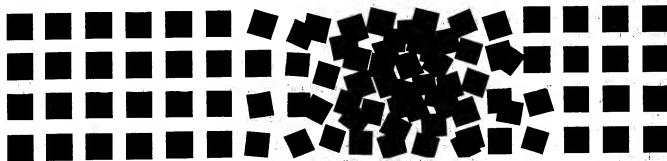
#2412

RC261
•C94
2001
PA/Bio

CANCER

*Principles & Practice
of Oncology*

6th Edition



www.LWWoncology.com



LIPPINCOTT WILLIAMS & WILKINS

A Wolters Kluwer Company

Philadelphia • Baltimore • New York • London
Buenos Aires • Hong Kong • Sydney • Tokyo

Acquisitions Editor: Stuart Freeman

Developmental Editors: Susan Rhyner, Managing Editor; Anne Snyder, Senior

Developmental Editor: and Stephanie Harris, Associate Developmental Editor

Supervising Editor: Toni Ann Scaramuzzo

Production Editors: Kim Langford and Shannon Garza, Silverchair Science + Communications

Manufacturing Manager: Tim Reynolds

Compositor: Silverchair Science + Communications

Printer: Quebecor World, Taunton, MA

6th edition

© 2001 by LIPPINCOTT WILLIAMS & WILKINS

530 Walnut Street

Philadelphia, PA 19106 USA

LWW.com

Copyright © 1993, 1989, 1985, 1982 by J.B. Lippincott Company. All rights reserved. This book is protected by copyright. No part of this book may be reproduced in any form or by any means, including photocopying, or utilized by any information storage and retrieval system without written permission from the copyright owner, except for brief quotations embodied in critical articles and reviews. Materials appearing in this book prepared by individuals as part of their official duties as U.S. government employees are not covered by the above-mentioned copyright.

Printed in the USA

Library of Congress Cataloging-in-Publication Data

Library of Congress Control Number: 89-649-721

Cancer: principles and practice of oncology [edited by] Vincent T. DeVita, Jr., Samuel Hellman, Steven A. Rosenberg; 319 contributors.—6th

ISSN 0892-0567

ISBN 0-781-72229-2

Care has been taken to confirm the accuracy of the information presented and to describe generally accepted practices. However, the authors, editors, and publisher are not responsible for errors or omissions or for any consequences from application of the information in this book and make no warranty, expressed or implied, with respect to the currency, completeness, or accuracy of the contents of the publication. Application of this information in a particular situation remains the professional responsibility of the practitioner.

The authors, editors, and publisher have exerted every effort to ensure that drug selection and dosage set forth in this text are in accordance with current recommendations and practice at the time of publication. However, in view of ongoing research, changes in government regulations, and the constant flow of information relating to drug therapy and drug reactions, the reader is urged to check the package insert for each drug for any change in indications and dosage and for added warnings and precautions. This is particularly important when the recommended agent is a new or infrequently employed drug.

Some drugs and medical devices presented in this publication have Food and Drug Administration (FDA) clearance for limited use in restricted research settings. It is the responsibility of health care providers to ascertain the FDA status of each drug or device planned for use in their clinical practice.

SECTION 3

OLIVER MICHAEL COLVIN

Antitumor Alkylating Agents

HISTORY OF THE ALKYLATING AGENTS

A nitrogen mustard alkylating agent was the first nonhormonal chemical that demonstrated significant clinical antitumor activity. The clinical evaluation of nitrogen mustards as antitumor agents evolved from the observed clinical effects of sulfur mustard gas used as a weapon in World War I. This gas was used because of its vesicant effect on the skin and mucous membranes, especially the eyes and respiratory tract.¹ However, in addition to this deadly effect, depression of the hematopoietic and lymphoid systems was observed in victims and experimental animals.² These observations led to further studies that used the less volatile nitrogen mustards (Fig. 19.3-1). Studies published in 1946 demonstrated regression of tumors, especially lymphomas³⁻⁵ and led to the introduction of the compound nitrogen mustard (mechlorethamine, Mustargen) into clinical practice. Subsequently, less toxic and more clinically effective nitrogen mustard derivatives and other types of alkylating agents have been developed.

CHEMISTRY AND CYTOTOXICITY OF ALKYLATING AGENTS

The alkylating agents react with (or "alkylate") many electron-rich atoms in cells to form covalent bonds. The most important reactions with regard to their antitumor activities are reactions with DNA bases. Some alkylating agents are monofunctional and react with only one strand of DNA. Others are bifunctional and react with an atom on each of the two strands of DNA to produce a "cross-link" that covalently links the two strands of the DNA double helix. Unless repaired, this lesion will prevent the cell from replicating effectively. The lethality of the monofunctional alkylating agents results from the recognition of the DNA lesion by the cell and the response of the cell to that lesion. Analogous cellular reactions may occur to the interstrand cross-links, but such reactions have not been definitively established.

CLASSES OF ALKYLATING AGENTS AND THEIR PROPERTIES

NITROGEN MUSTARDS

Mustargen

Mustargen is currently used in the MOPP [Mustargen, vincristine (Oncovin), procarbazine, prednisone] regimen for the treatment of Hodgkin's disease⁶ but rarely for other purposes. The other nitrogen mustards in significant clinical use are cyclophosphamide, ifosfamide, melphalan, and chlorambucil (Fig. 19.3-2). All these compounds produce cytotoxicity by forming covalent interstrand cross-links in DNA (as shown in Fig. 19.3-3 for Mustargen). The nitrogen mustard cross-link has been demonstrated to occur in the G-X-C/C-Y-G configura-

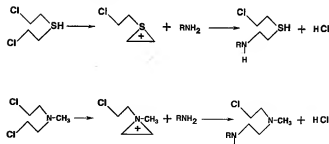


FIGURE 19.3-1. Structures and alkylation mechanisms for sulfur mustard and the nitrogen mustard, Mustargen (mechlorethamine).

tion,⁷ as opposed to the G-C/C-G cross-link that had previously been predicted.⁸ The formation of the G-X-C/C-X-G cross-link has been postulated to occur on the basis of the greater frequency of approximation of the N7 atoms of the two guanylates in the G-X-C/C-X-G configuration, as opposed to the G-C/C-G configuration.⁹

Mustargen is available only as an intravenous preparation that can also be used topically for cutaneous malignancies. In the MOPP regimen, Mustargen is used at a dose of 6 mg/m² on days 1 and 8 of the monthly schedule. Toxicities unique to the agent are topical irritation and pain on injection if given too rapidly. The clearance of the drug is very rapid, but pharmacokinetics have not been performed with modern techniques.

Cyclophosphamide

The most frequently used alkylating agent, cyclophosphamide, is used for the treatment of breast cancer in combination with doxorubicin (Adriamycin)¹⁰ or with methotrexate and 5-fluorouracil¹¹ and for the treatment of lymphomas,^{12,13} childhood tumors,^{14,15} and many solid tumors.¹⁶ High doses of cyclophosphamide are frequently used in conjunction with bone marrow transplantation¹⁷⁻¹⁹ and for the treatment of autoimmune diseases.^{20,21}

Cyclophosphamide¹ is inactive *in vitro* and is metabolized by P-450 enzymes in the liver to active species, as shown in Figure 19.3-4. The initial product is 4-hydroxycyclophosphamide (4HC), which is released from the liver into the circulation.²² This compound is in equilibrium with an open-ring tautomer, aldophosphamide. Aldophosphamide spontaneously eliminates acrolein to produce phosphoramidate mustard,²³ which is an active bifunctional alkylating species.²⁴ Phosphora-

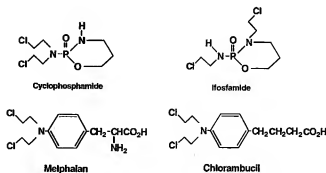


FIGURE 19.3-2. Nitrogen mustards in frequent clinical use.

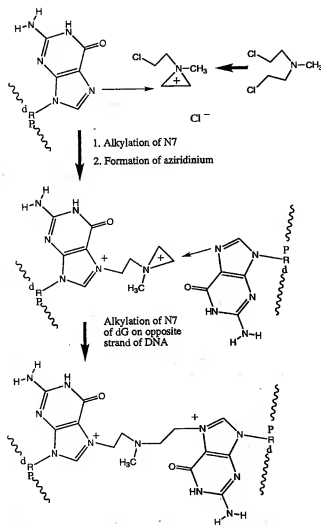


FIGURE 19.3-3. Alkylation of DNA and formation of interstrand cross-link by nitrogen mustard.

midate mustard is zwitterionic at physiologic pH²⁵ and enters cells poorly. 4-HC-aldophosphamide is not charged and enters cells readily. While phosphoramide mustard is toxic to cells *in vitro* at concentrations of 100 μ M and higher, 4-HC is cytotoxic in the range of 10 μ M.²⁶ Thus, 4-HC-aldophosphamide serves as an efficient delivery system for phosphoramide mustard, which has been demonstrated to produce an interstrand DNA cross-link analogous to the cross-link produced by mechlorethamine.⁷ Recent studies by Shulman-Roskes et al.²⁷ have demonstrated that phosphoramide mustard readily eliminates chloroethylaziridine,²⁷ which probably also plays a role in the cross-linking of DNA in cells exposed to 4-HC.

As shown in Figure 19.3.4, 4-HC is a substrate for the enzyme aldehyde dehydrogenase.²⁸ In cells that contain this enzyme, the bulk of the 4-HC is oxidized to carboxyphosphamide, which is not an active alkylating agent. Consequently, cells with high aldehyde dehydrogenase (ALDH) content are resistant to the metabolites of cyclophosphamide.^{29,30} Early hematopoietic stem cells and megakaryocytes contain high levels, as do the epithelial stem cells in the small intestine and mucous membranes.^{30,31} These observations explain why cyclo-

phosphamide administration produces a shorter period of hematopoietic depression,³² is relatively sparing of platelets, and is associated with less gastrointestinal toxicity and mucositis than other alkylating agents.³³

4-HC is too unstable to be used as a reagent, but the compound 4-hydroperoxycyclophosphamide (see Fig. 19.3-4) is spontaneously converted in aqueous solution to 4-HC and can be used for *in vitro* studies of cell sensitivity.^{34,35} This compound has also been used for the *in vitro* treatment of autologous bone marrow to reduce the number of tumor cells returned to the patient.³⁶

Cyclophosphamide is available as tablets for oral administration or as an intravenous preparation. The drug is used at a variety of doses and schedules. Oral administration is particularly used for autoimmune diseases at a daily dose of approximately 100 mg. Because of its rapid absorption and high bioavailability, even very high doses can be given orally, but high intermittent doses are usually given intravenously. In moderate-dose combination chemotherapy, doses of cyclophosphamide in the range of 750 mg are usually used. For high-dose therapy in conjunction with hematopoietic cell transplantation, doses of up to 50 mg/kg for 2 or 4 days in combination with other agents are used.

The bulk (nearly 70%) of a dose of cyclophosphamide is excreted in the urine as the inactive carboxyphosphamide.^{37,38} At high doses (approximately 50 mg/kg), plasma concentrations of up to 400 μ M of cyclophosphamide are achieved,³⁸ and clearance depends on the renal clearance and the rate of microsomal metabolism in the liver. With improved and more facile techniques to measure 4-HC concentrations accurately, the clinical pharmacology of cyclophosphamide and this critical transport intermediate are being more carefully defined. Studies in patients receiving high-dose therapy have demonstrated considerable variation in the rates of clearance of cyclophosphamide between patients, with consequent differences in the peak concentrations (1 to 15 μ M) and total exposure of the patient to 4-HC (60 to 140 μ M.hours).^{39,40} The total exposure to 4-HC is probably the major determinant of therapeutic effect. Currently, several programs are evaluating dose adjustment regimens based on the initial pharmacokinetics of cyclophosphamide and 4-HC. While it is known that substantial concentrations of phosphoramide mustard are present in plasma (up to 10 μ M after 60 mg/kg of cyclophosphamide³⁹), this concentration is well below the concentrations needed for *in vitro* cytotoxicity of phosphoramide mustard.³⁶

A unique toxicity of cyclophosphamide and other oxazophosphorines is a characteristic hemorrhagic cystitis^{41,42} due to irritation of the bladder mucosa from urinary metabolites. Acrolein has been identified as the metabolite most responsible for this effect,⁴³ but phosphoramide mustard and chloroacetaldehyde may contribute to this toxicity. Careful hydration and emptying of the bladder are crucial to avoiding this toxicity, which has produced massive and even fatal hemorrhage. Another toxicity that has been associated with cyclophosphamide is an antidiuretic effect, especially at high doses.⁴⁴ This effect may produce marked fluid retention and electrolyte abnormalities, particularly low sodium, and seizures and fatalities have been seen.⁴⁵ It is important to avoid low-sodium-containing fluids after high-dose cyclophosphamide, and the fluid retention syndrome has been treated with furosemide to promote free water clearance.⁴⁶ The most severe dose-limiting toxicity of cyclophosphamide is a fulminant cardiac toxicity,⁴⁷ which is often fatal when seen clinically.

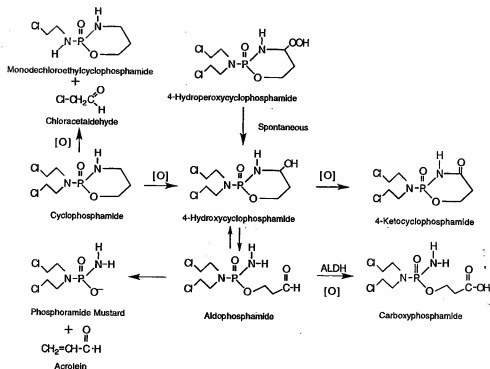


FIGURE 19.3-4. Metabolism of cyclophosphamide.

cally. This toxicity is seen only after the high doses used in bone marrow transplantation. It was initially seen in patients receiving 60 mg/kg/d of cyclophosphamide for 4 days, and the incidence has decreased since lower doses have been used. The syndrome usually presents with severe cardiac failure, beginning approximately 10 days after drug administration, with a dilated heart and low electrocardiogram voltage. There is a characteristic pathologic picture of edema, interstitial hemorrhage, and cardiac necrosis.⁴⁷

Ifosfamide

Ifosfamide is a structural isomer of cyclophosphamide that is often used in the treatment of sarcomas and pediatric tumors (see Fig. 19.3-2). There is more chloroethyl side chain oxidation of ifosfamide (up to 50%) than of cyclophosphamide (<10%), and the degree of such metabolism is more variable than with cyclophosphamide.⁴⁸ Oxidation of the chloroethyl groups produces chloroacetaldehyde, which is probably responsible for the neurotoxicity⁴⁹ and renal toxicity⁵⁰ that have been seen with ifosfamide therapy. Since the oxidation of a chloroethyl side chain produces a much less toxic monofunctional agent, higher doses of ifosfamide than cyclophosphamide must be used clinically. The studies of the clinical pharmacology of ifosfamide have been more limited than those of cyclophosphamide but have demonstrated large intrapatient variability in the pharmacokinetics and metabolism of the agent during repeated administrations.^{51,52}

Melphalan

Melphalan is now used principally for the treatment of multiple myeloma,⁵³ for high-dose myeloblastic therapy in conjunction with bone marrow transplantation,⁵⁴ and for the isolated

limb perfusion of localized tumors,⁵⁵ especially malignant melanoma and sarcomas (see Fig. 19.3-2). Melphalan is an amino acid analogue and is actively transported into cells by amino acid transport systems.^{56,57} It has been demonstrated that cellular uptake⁵⁸ and transport into the central nervous system (CNS)⁵⁹ of melphalan can be modulated by the amino acid content in the extracellular fluid.

Melphalan is available both as tablets and as an intravenous preparation. For the treatment of multiple myeloma, melphalan is usually used orally at a dose of 0.25 mg/kg for 4 days, with prednisone on the same schedule every 4 to 6 weeks. At these doses, peak plasma concentrations of 0.625 μ M are found, but absorption is variable.⁶⁰ For bone marrow transplantation, doses of melphalan of 100 to 140 mg/m² are used.⁵¹ At these doses, peak concentrations of melphalan of 40 to 50 μ M are reached.^{61,62}

Chlorambucil

Chlorambucil is used for the treatment of B-cell chronic lymphocytic leukemia⁶³ and lymphomas⁶⁴ and for the immunosuppressive therapy of autoimmune diseases.⁶⁵ It is administered orally and is well tolerated when given either by daily administration or intermittent high-pulse doses.⁶⁴ Chlorambucil is well tolerated by most patients and can be used successfully for patients who have severe nausea and vomiting with cyclophosphamide or melphalan.

Chlorambucil is available only in an oral formulation. For chronic leukemia and immunosuppression, daily doses of 3 to 6 mg are given for a number of weeks, or 12 mg/m² may be given monthly. Pulsed dose pulse chlorambucil for lymphoma is given orally at a dose of 16 mg/m² daily for 5 consecutive days each month.⁶⁴ Chlorambucil is metabolized to a less active derivative—phenylacetic acid mustard—and the clinical pharmacology of chlorambucil is very similar to that of melphalan.⁶⁶

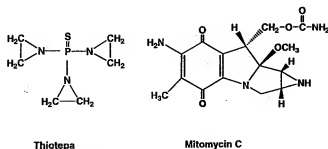


FIGURE 19.3-5. Aziridine agents.

AZIRIDINES AND EPOXIDES

The aziridine agents are related to the nitrogen mustards but contain uncharged aziridine rings that are less reactive than the aziridinium rings formed by most of the nitrogen mustards. The two aziridine agents that are frequently used clinically are thiotepea and mitomycin C (Fig. 19.3-5). The diepoxide dianhydrogalactitol reacts with DNA in a similar fashion to the aziridines but has been succeeded in clinical use by dibromodulcitol, which spontaneously generates dianhydrogalactitol *in situ* (Fig. 19.3-6).

Thiotepea

Thiotepea is now used most frequently in combination with other alkylating agents in high-dose therapy with stem cell support.^{86,87} Thiotepea has been demonstrated to react with the N7 position of guanylic acid in DNA⁸⁸ and to cross-link DNA,⁸⁹ indicating that it is acting similarly to the nitrogen mustards. Thiotepea is desulfurated by cytochrome P-450 enzymes⁷⁰ to produce tepa. Tepa is less toxic than thiotepea and has been demonstrated to produce alkali-labile sites in DNA, rather than cross-links.⁸⁹ These findings suggest that tepa reacts differently from thiotepea and produces monofunctional alkylation of DNA.

In combination with cyclophosphamide for high-dose therapy, thiotepea has been given as a continuous infusion for 4 days, at a daily dose of 200 mg/m². Under these conditions, steady-state levels of 2 to 6 μM of thiotepea are rapidly achieved.⁷¹ Thiotepea is also used at a dose of 900 mg/m² in combination with high-dose cyclophosphamide and cisplatin.⁷²

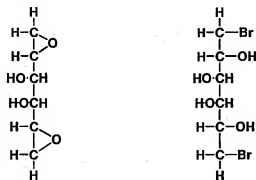


FIGURE 19.3-6. Structures of dianhydrogalactitol and its pro-drug, dibromodulcitol.

Mitomycin C

Mitomycin C is an antibiotic extracted from a *Streptomyces* species and is used for the treatment of breast cancer,⁷³ esophageal cancer,⁷⁴ and gastrointestinal tumors.⁷⁵ As seen in Figure 19.3-5, this compound contains an aziridine ring. Particularly under hypoxic conditions, mitomycin C is reduced, with activation of the C1 position of the aziridine ring. This carbon then reacts in the minor groove with the extracyclic N2 amino group of a guanylic acid,^{76,77} positioning the 10 carbon of the carbamate moiety to react with the N2 of a guanylic acid residue in an adjacent base pair in the complementary DNA strand. Mitomycin C and its reduced metabolites can also produce intrastrand guanylic acid–guanylic acid cross-links that produce bending of the DNA.⁷⁸

In combination regimens, mitomycin C is given at doses of 10 to 15 mg/m² every 4 to 6 weeks. After a dose of 15 mg/m², peak plasma concentrations of 3 μM are seen.⁷⁹

Dianhydrogalactitol

Dianhydrogalactitol (see Fig. 19.3-6) is a hexitol derivative that contains two epoxide groups and cross-links DNA through the N7 atoms of guanylic acid,⁸⁰ presumably through the nucleophilic attack of the N7 atoms on the strained-ring epoxide groups. This compound was evaluated in clinical trials and demonstrated modest antitumor activity.^{81,82} However, the structurally related dibromodulcitol (see Fig. 19.3-6) has demonstrated more antitumor activity^{83,84} and is still being used in combination chemotherapy of breast cancer, cervical cancer, and brain tumors. Dibromodulcitol is hydrolyzed to dianhydrogalactitol, and its better antitumor activity is presumably due to more effective localization of the reactive agent in tumor cells.⁸⁵ Dibromodulcitol is usually administered at a dose of 1 g/m², which produces a maximum plasma concentration of approximately 50 μM.⁸⁵

ALKYL SULFONATES: BUSULFAN

Busulfan (Mylceran), other alkyl sulfonates, and the related sulfamates react with DNA by a direct displacement reaction (as shown in Fig. 19.3-7). Busulfan has been demonstrated to cross-link DNA,⁸⁷ but the structure of the cross-link has not been established. A chemically related agent, hepsulfan, with seven methylene units between the reactive groups, has been demonstrated to form a DNA G-X-C/C-X-G interstrand cross-link analogous to those formed by the nitrogen mustards.⁸⁸ Haddow and Timmis⁸⁹ reported in 1953 that busulfan was active against chronic myelogenous leukemia. Busulfan was for many years the principal agent used to treat this disease before being replaced by the use of hydroxyurea⁹⁰ and interferon-α,⁹¹ both of which have proved to be more effective than busulfan. The most frequent use of busulfan in cancer therapy today is in high-dose therapy for many tumors including chronic myelogenous leukemia, in conjunction with bone marrow or stem cell transplantation. For this application, high doses of busulfan are combined with cyclophosphamide, total body irradiation, or other agents.^{18,92-94} The effectiveness of busulfan for this purpose is undoubtedly related to its marked myeloablative properties,⁹⁵ the mechanisms of which are not understood.

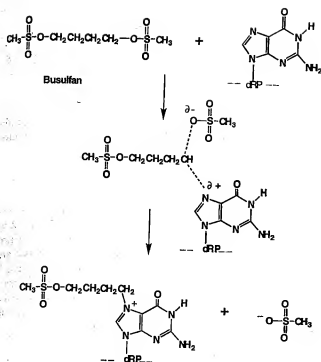
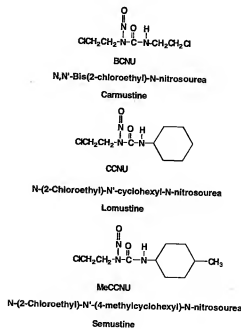


FIGURE 19.3-7. Alkylation of guanylate in DNA by busulfan through S₂ alkylation. A second displacement reaction with the N7 of a guanylate in the complementary strand creates a G-X-C/C-X-G interstrand cross-link.

Until recently, busulfan was available only as an oral preparation, but intravenous preparations are now available. For hematopoietic transplantation, busulfan is usually given as 1 mg/kg every 6 hours for 4 days, for a total dose of 16 mg/kg. Peak concentrations of busulfan after each dose are approximately 10 μ M.⁹⁶ High doses of busulfan have been associated with venoocclusive disease of the liver. This syndrome consists of hepatomegaly, jaundice, ascites, and hepatic failure with a high mortality rate.⁹⁷ Grochow et al.⁹⁶ have demonstrated that pharmacokinetic monitoring and dose adjustment of the busulfan can markedly reduce the incidence of venoocclusive disease.

NITROSOUREAS

The members of the nitrosourea group of therapeutic alkylating agents are related to the alkylnitrosoamines and similar compounds that have long been known to be carcinogenic. Methylnitrosoguanidine and methylnitrosourea are monofunctional alkylating agents and were found to have modest antitumor activity.^{98,99} Montgomery¹⁰⁰ and others^{101,102} evaluated a number of analogues of these compounds and demonstrated remarkable antitumor effects of bischloroethylnitrosourea (BCNU; Fig. 19.3-8) against mouse tumors, and particularly against intracerebral tumors, which had been refractory to most agents because of the blood-brain barrier.¹⁰⁰⁻¹⁰² BCNU was found to produce interstrand cross-linking of DNA,¹⁰³ which has been demonstrated to occur through the spontaneous generation of a chloroethyl diazonium species¹⁰⁴ and the series of reactions illustrated in Figure 19.3-9.¹⁰⁵ As illustrated, this interstrand cross-link occurs between



N'-(4-Amino-2-methyl-5-pyrimidinyl)methyl-N-(2-Chloroethyl)-N-nitrosourea

Nimustine

FIGURE 19.3-8. Nitrosoureas.

a guanylate in DNA and the base-paired cytidylate in the other strand of the DNA.¹⁰⁶

Bischloroethylnitrosourea

BCNU (carmustine; see Fig. 19.3-8) demonstrated activity against brain tumors clinically¹⁰⁷ and has continued to be used in the treatment of gliomas and other brain tumors. BCNU has also been used in the treatment of multiple myeloma¹⁰⁸ and in high-dose therapy in conjunction with bone marrow and stem cell transplantation.¹⁰⁹ BCNU can also be administered to brain tumors by direct injection¹¹⁰ and by the implantation of biodegradable polymers containing BCNU into the brain.¹¹¹

Cyclohexylchloroethylnitrosourea

Cyclohexylchloroethylnitrosourea (CCNU, lomustine; see Fig. 19.3-8) is a more lipid-soluble nitrosourea. It is administered orally and is used in the treatment of brain tumors.^{112,113}

Methylcyclohexylchloroethylnitrosourea

Methylcyclohexylchloroethylnitrosourea (semustine; see Fig. 19.3-8) is an oral investigational drug that has been used in the treatment of gastrointestinal tumors.¹¹⁴

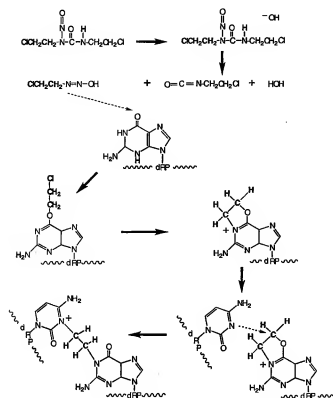


FIGURE 19.3-9. Reaction of BCNU with DNA to produce a G-C interstrand cross-link.

N'-[(4-amino-2-methyl-5-pyrimidinyl)methyl]-N-(2-chloroethyl)-N-nitrosourea

N'-[(4-amino-2-methyl-5-pyrimidinyl)methyl]-N-(2-chloroethyl)-N-nitrosourea (nimustine; see Fig. 19.3-8) is more water-soluble than the other chloroethylnitrosoureas and has been used for the treatment of CNS tumors by the intraarterial¹¹⁵ and intrathecal routes.¹¹⁶

Clinical Pharmacology

As a single agent, BCNU is usually used in a dose of 125 to 200 mg/m² every 6 to 8 weeks. In combination with doxorubicin for multiple myeloma, a dose of 30 mg/m² every 3 to 4 weeks has been used.¹¹⁷ After doses in the range of 100 mg/m², peak plasma concentrations are in the range of 5 µM.¹¹⁸ For high-dose therapy of breast cancer, BCNU is given at a dose of 600 mg/m² in combination with cyclophosphamide and cisplatin.¹¹⁹ After this dose of BCNU, the peak plasma levels of BCNU have been shown to be approximately 5 µM.¹²⁰ Phenobarbital has been demonstrated to increase the clearance of BCNU¹²¹ and to decrease the toxic and therapeutic effects. CCNU is administered in doses similar to those of BCNU. The parent CCNU has not been detected, but the peak concentrations of the ring hydroxylated metabolites are approximately 3 µM after doses of 130 mg/m².¹²²

Specific Toxicities

Hematopoietic toxicity of the nitrosoureas is severe and is delayed, with the nadir of the granulocytes occurring approxi-

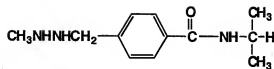


FIGURE 19.3-10. Procarbazine.

mately 5 to 6 weeks after administration.¹²³ This finding indicates that these agents selectively damage a very primitive hematopoietic precursor.

HYDRAZINE AND TRIAZINE DERIVATIVES

The hydrazine and triazine derivative compounds are analogous to the nitrosoureas in that they decompose spontaneously or are metabolized to produce an alkyl carbonium ion, which alkylates DNA. Hydrazine and its substituted analogues are known carcinogens¹²⁴ that inhibit gluconeogenesis in cells¹²⁵ and have been promoted as antitumor agents.¹²⁶ However, objective preclinical and clinical studies have not supported a significant antitumor effect^{127,128} for hydrazine analogues in general.

Procarbazine

Procarbazine is a phenylhydrazine derivative that was initially developed as an inhibitor of monoamine oxidase but was found to have significant antitumor activity in preclinical models and clinically (Fig. 19.3-10).¹²⁹ Procarbazine was one of the components of the first effective combination chemotherapy regimen, MOPP, for Hodgkin's disease.⁶ The agent is currently used for the treatment of Hodgkin's disease,^{5,130} and for the treatment of primary brain tumors.^{113,131} Procarbazine has been demonstrated to be metabolized to a DNA-methylating agent,¹³²⁻¹³⁴ which is most likely methylazoxypcarbazine.^{135,136} Since procarbazine is a monoamine oxidase inhibitor, patients can experience CNS depression,¹³⁷ or stimulation¹³⁸ and acute hypertension, especially after the ingestion of tyramine-rich foods.

Dacarbazine

Dacarbazine, or DTIC [(dimethyltriazene)imidazole-carboxamide], is a triazine derivative that is metabolized by microsomal N-demethylation, predominantly in the liver, to an intermediate that spontaneously decomposes to release a methyl diazonium ion that methylates DNA (Fig. 19.3-11).¹³⁹⁻¹⁴¹ Dacarbazine is used in the regimen of doxorubicin, bleomycin, vinblastine, and dacarbazine for the treatment of Hodgkin's disease^{130,142} and for the treatment of malignant melanoma.^{119,143}

Temozolomide

Temozolomide is a triazine analogue that spontaneously decomposes to produce a methyl diazonium ion, as illustrated in Figure 19.3-11.^{144,145} This compound may produce a more homogeneous distribution of the short-lived MITC [(methyltriazene)imidazole-carboxamide], which is spontaneously generated from temozolomide at all sites, than does dacarbazine, which is metabolized to MITC in the liver. The principal toxicities seen in phase I trials have been neutropenia and thrombocytopenia.

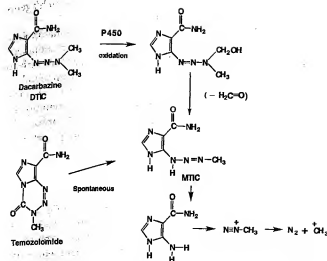


FIGURE 19.3-11. Generation of methyl diazonium from the triazene dacarbazine and temozolomide.

and tumor responses were seen in those trials^{146,147} in patients with glioma and melanoma. Phase II trials in patients with gliomas have shown response rates of 20% to 30%,^{148,149} but phase II trials in patients with sarcomas¹⁵⁰ and pancreatic cancer¹⁵¹ did not demonstrate significant responses.

These agents exert their toxicity predominantly through the methylation of the O^6 position of guanylic acid in DNA. Therefore, cells that contain significant O^6 -alkyltransferase or are deficient in mismatch repair will be resistant to them (as discussed in the section Mechanisms of Toxicity and Drug Resistance).

Procarbazine is an oral preparation and used in the MOPP regimen for Hodgkin's disease at a dose of 100 $\text{mg}/\text{m}^2/\text{d}$ for 14 days.¹⁴² Because of its complex metabolism, pharmacokinetic studies have been limited. Dacarbazine is an intravenous preparation and is used in the regimen of doxorubicin, bleomycin, vinblastine, dacarbazine for Hodgkin's disease at a dose of 375 $\text{mg}/\text{m}^2/\text{d}$ for 15 days.¹⁴² For the treatment of malignant melanoma, a dose of 200 to 250 $\text{mg}/\text{m}^2/\text{d}$ for 5 days is used and, at this dose, peak plasma concentrations of dacarbazine are approximately 30 μM .¹⁵² This agent has been used as a single agent with bone marrow transplantation at a dose of 2000 mg/m^2 .¹⁵³ At this dose, the maximum plasma concentration of dacarbazine was 800 μM .¹⁵³ Temozolomide is usually given orally at 150 to 250 $\text{mg}/\text{m}^2/\text{d}$ for 5 days. Reid et al.¹⁵⁴ measured peak concentrations of MTIC of 0.5 to 5 μM after administration of these doses of temozolomide.¹⁵⁴ Baker et al.¹⁵⁵ studied the pharmacokinetics of ¹⁴C-labeled temozolomide and found peak concentrations of temozolomide of approximately 30 μM and peak concentrations of MTIC of approximately 1 μM .

MECHANISMS OF TOXICITY AND DRUG RESISTANCE

REACTION WITH CELLULAR MOLECULES

The alkylating agents are potent electrophiles and react with many electron-rich molecules within the cell to be inactivated.

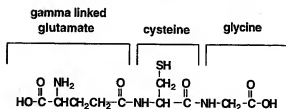


FIGURE 19.3-12. Structure of glutathione.

The principal such molecule is glutathione (GSH), a tripeptide with a free cysteine sulfhydryl that is present at millimolar concentrations in cells (Fig. 19.3-12). This small nucleophile is known to react with and inactivate virtually all the therapeutic alkylating agents, and a correlation between elevated cellular GSH concentrations and resistance to nitrogen mustards has been demonstrated.^{156,157} The GSH S-transferase enzymes catalyze the conjugation of GSH with electrophiles, and increased activity of this class of enzymes enhances GSH-mediated resistance.¹⁵⁸⁻¹⁶⁰ The GSH conjugates of specific alkylating agents have been characterized,¹⁶¹⁻¹⁶³ and the specific isoenzymes of GST that catalyze their formation have been characterized.¹⁶⁴⁻¹⁶⁸

Buthionine sulfoximine is an inhibitor of gamma-glutamylcysteine synthetase, the rate-limiting enzyme in the GSH synthesis pathway, and decreases the GSH concentration in cells.¹⁶⁹ Exposure to this compound sensitizes both normal and tumor cells to alkylating agents.^{166,170,171} In a phase I clinical trial, buthionine sulfoximine has been shown to increase the hematologic toxicity of melphalan¹⁷² and is currently in further clinical trials to determine whether this agent can increase the clinical antitumor efficacy of melphalan.

Cells can also be sensitized to alkylating agents by exposure to inhibitors of GSH S-transferase,^{173,174} and a clinical trial of the GSH S-transferase inhibitor sulfasalazine with melphalan demonstrated increased nausea and vomiting but no increase in hematopoietic toxicity.¹⁷⁵ The membrane transporter multidrug resistance protein is known to mediate the efflux of GSH conjugates from the cell,¹⁷⁶ and Barnouin et al.¹⁷⁷ have demonstrated that this system can transport the GSH conjugates of chlorambucil and melphalan from cells. The observations suggest that modulation of these systems could enhance the efficacy of alkylating agents.

Kelley et al.¹⁷⁸ demonstrated that transfection of metallothionein into cells produced increased resistance to chlorambucil and melphalan. Subsequently, Yu et al.¹⁷⁹ have demonstrated that the thiol groups of metallothionein will bind melphalan and phosphoramide mustard.¹⁸⁰ It has also been demonstrated that exposure of cells to zinc will increase metallothionein concentration in the cell and increase resistance of the cells to melphalan, doxorubicin, and cisplatin.¹⁸¹

ENHANCED DNA REPAIR: O^6 ALKYLATION

Another mechanism of cellular resistance to alkylating agents is repair of the DNA damage that the agents produce. The most defined mechanism of cellular repair of alkylating agent damage is that of the enzyme O^6 -alkylguanine-alkyltransferase. As illustrated in Figure 19.3-13, this enzyme can remove an alkyl group from the O^6 position of guanine, and the alkylated enzyme is then rapidly degraded.¹⁸² This mechanism has been shown to be effective in protecting normal and tumor cells

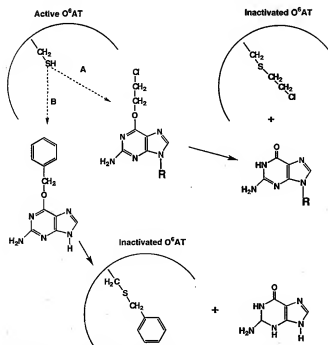


FIGURE 19.3-13. Interactions of O^6 -alkylguanine-DNA alkyltransferase. Pathway A: Repair of O^6 alkylation by O^6 -AT. Pathway B: Inactivation of O^6 -AT by benzylguanine.

from the carcinogenic and toxic effects of DNA methylating agents, such as temozolomide and procarbazine.¹⁸⁸ Erickson et al.¹⁸⁴ demonstrated that this enzyme would also remove the 6-chloroethyl lesion produced by the alkylation of guanine by the chloroethylnitrosoureas and produce resistance to these compounds, and this observation has been confirmed and extended.¹⁸⁵

It has been shown that such compounds as O^6 -benzylguanine will be acted on by O^6 -alkylguanine-DNA alkyltransferase (see Fig. 19.3-13) to remove the benzyl group¹⁸⁶ and that the enzyme will be rapidly degraded and depleted. Such compounds have been demonstrated to reverse tumor resistance due to O^6 -AT to the O^6 alkylating agents *in vitro* and *in*

vivo,^{187,188} and clinical trials of the combination of such agents and O^6 -methylguanine are currently in progress.^{189,190}

However, inhibitors of O^6 -AT enhance the hematopoietic toxicity of O^6 alkylating therapeutic agents. Hematopoietic stem cells have been successfully transfected with O^6 -AT variants that are resistant to O^6 -benzylguanine and related compounds.¹⁹¹ The hematopoietic systems of animals populated with these cells are resistant to the combination of O^6 -benzylguanine and BCNU,¹⁹² and clinical trials of this approach to improve the efficacy of chloroethylnitrosoureas and methylating agents are planned.

CROSS-LINK REPAIR

The use of alkaline elution and other techniques (Fig. 19.3-14) has demonstrated that DNA interstrand cross-links produced by nitrogen mustards can be removed in bacteria¹⁹³ and mammalian cells.¹⁹⁴ The mechanism of such repair has not been elucidated, but nucleotide excision repair¹⁹⁵ and poly(adenosine diphosphate-ribose) polymerase¹⁹⁶ appear to play a role.

Caffeine and related compounds have been demonstrated to enhance the cytotoxicity of nitrogen mustard.¹⁹⁷ This effect was associated with abrogation of G_2 arrest. O'Connor et al.^{198,199} demonstrated that the G_2 arrest associated with nitrogen mustard resistance was associated with decreased activity of cdc2 kinase in the resistant cells. Caffeine has also been shown to inhibit nucleotide excision repair by binding to the subunit that recognizes the damage and helps to mediate this repair activity.²⁰⁰ Elevated Bcl-2 has also been associated with nitrogen mustard resistance.²⁰¹

A medulloblastoma cell line has been demonstrated to be resistant to activated cyclophosphamide (4-hydroperoxycyclophosphamide) on the basis of increased removal of DNA interstrand cross-links.^{34,202} This cell does not appear to repair cross-links produced by BCNU and busulfan, indicating that the recognition of the nitrogen mustard cross-link is fairly specific.

IN VIVO RESISTANCE

Kobayashi et al.²⁰³ and St. Croix et al.²⁰⁴ have described resistance to alkylating agents and other antitumor agents that

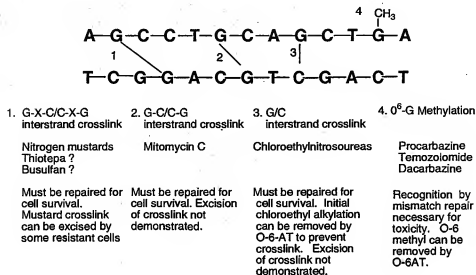


FIGURE 19.3-14. DNA lesions produced by alkylating agents.

associated with aggregation of tumor cells. This resistance is present when the tumor cells are growing *in vivo* or in three-dimensional *in vitro* culture with adherence between the cells but is not present when the cells are dispersed in two-dimensional culture. This type of resistance has also been associated with increased metastatic potential.²⁰⁵

COMMON TOXICITIES

Toxicities that are associated with specific alkylating agents are described in the discussions of the individual agents. The toxicities common to the alkylating agents as a class are described here.

HEMATOPOIETIC TOXICITY

The usual dose-limiting toxicity for an alkylating agent is hematopoietic toxicity. As described, cyclophosphamide usually produces a relatively rapid nadir of the granulocytes, with recovery within 3 weeks after a single dose or short course.^{32,33,206} Cyclophosphamide is also relatively platelet-sparing. The reason for the relative hematopoietic sparing properties of cyclophosphamide is the high concentrations of the enzyme aldehyde dehydrogenase in hematopoietic stem cells and megakaryocytes.^{30,31}

The nitrosoureas produce an unusual delayed hematopoietic toxicity, with nadirs of both granulocytes and platelets at 5 to 6 weeks after administration.¹²³ Severe granulocytopenia and thrombocytopenia are also characteristic of busulfan.²⁰⁷ An interesting characteristic of busulfan is its relative sparing of lymphocytes. The different hematopoietic effects of alkylating agents, except for the characteristics of cyclophosphamide, are not explained but suggest significant differences in selectivity of the agents for hematopoietic precursors.

GASTROINTESTINAL TOXICITY

The alkylating agents frequently produce nausea and vomiting, although this effect is usually not as severe as with the platinum agents. Cyclophosphamide produces severe nausea and vomiting in some patients, but these patients usually tolerate chlorambucil, which is clinically less emetogenic. The nausea and vomiting produced by alkylating agents are known to be mediated significantly through the CNS.^{208,209} With the higher doses of alkylating agents used in bone marrow transplantation, increased nausea and vomiting are seen but can usually be controlled by corticosteroids and the newer antiemetic antiemetics.²¹⁰⁻²¹² The alkylating agents can cause significant toxicity to the gastrointestinal mucosa and produce mucositis, stomatitis, and diarrhea, especially with the high doses of melphalan and thiopeta used in bone marrow transplantation.²¹³

GNADAL TOXICITY

The alkylating agents can produce significant gonadal toxicity. The characteristic testicular lesion in men is depletion of germ cells without damage to the Sertoli cells, which was first described with nitrogen mustard in 1948.²¹⁴ This lesion is also seen, often in association with oligospermia or aspermia, after treatment with other alkylating agents.^{215,216} Spermatogenic dysfunction is reversible in some patients.^{217,218}

Women treated with alkylating agents may develop amenorrhea associated with a marked decrease in ovarian follicles.^{215,219,220} This complication and its irreversibility increase with the age of the woman.²²¹

PULMONARY TOXICITY

Interstitial pneumonitis and fibrosis were initially reported as a consequence of busulfan therapy²²² but have subsequently been reported to occur after therapy with melphalan,²²³ chlorambucil,²²⁴ cyclophosphamide,^{225,226} mitomycin C,²²⁷ and BCNU.^{228,229} The clinical manifestations of this toxicity are dyspnea and a nonproductive cough, which can progress to cyanosis, pulmonary insufficiency, and death. The syndrome has particularly been associated in frequency and severity with high doses of BCNU.^{230,231} The greater pulmonary toxicity of BCNU may be due to the spontaneous decomposition of BCNU, which produces chloroethyl isocyanate in addition to the alkylating chloroethyl diazonium moiety described.²³² Chloroethyl isocyanate is an analogue of methyl isocyanate, a known pulmonary toxin that produced many deaths when released in an industrial accident in Bhopal, India.²³³

ALOPECIA

Alopecia from chemotherapy was first described after administration of dimethylmyeleran, an analogue of busulfan.²³⁴ The alkylating agents now most associated with alopecia are cyclophosphamide and ifosfamide. Feil and Lamoureux²³⁵ examined the alopecia-producing effects of metabolites and analogues of cyclophosphamide and proposed that the alopecic effect was due to the facile entry of a lipophilic metabolite (now known to be 4-HC) into the hair follicles. This hypothesis is consistent with the fact that vincristine, doxorubicin, and the taxanes, all associated with alopecia, are fairly lipophilic.

TERATOGENICITY

All the therapeutically used alkylating agents are teratogenic in animal studies.²³⁶⁻²³⁹ A review of the literature in 1968 found that 4 of 25 children born to mothers who received alkylating agents during the first trimester of pregnancy had fetal malformations.²⁴⁰ On the basis of the limited information available, women treated with an alkylating agent during the first trimester of pregnancy may have a risk as high as 15% of having a malformed infant. Administration of alkylating agents during the second and third trimesters has not been associated with increased fetal malformations.^{241,242} More recent reviews support the lack of malformations produced by treatment during the second and third trimesters,^{243,244} and one review cites 19 women treated during the first trimester with no infant malformations.²⁴⁴

CARCINOGENESIS

In the 1970s, there were reports of acute leukemia occurring in patients who had been treated with alkylating agents,²⁴⁵⁻²⁴⁹ and subsequent experience has confirmed the occurrence of this complication. The incidence of leukemia is difficult to estimate because of the variety of agents, doses, and combinations used but is probably approximately 5%. In one group of 12 ovarian cancer patients receiving a high dose of melphalan, 4 devel-

oped acute leukemia.²⁴⁸ In one report, the incidence of leukemia was found to be higher after melphalan treatment than after cyclophosphamide therapy.²⁵⁰ This observation may be related to the stem cell-sparing properties of cyclophosphamide.³⁰ An increased frequency of solid tumors also occurs after alkylating agent therapy.^{251,252}

IMMUNOSUPPRESSION

In 1921, Hektoen and Corper²⁵³ reported an inhibitory effect of sulfur mustard on antibody production. While all the alkylating agents produce some degree of immunosuppression, cyclophosphamide is the most immunosuppressive.²⁵⁴ Cyclophosphamide and chlorambucil are the alkylating agents most commonly used for the treatment of autoimmune diseases.²⁵⁵⁻²⁵⁹

Selective inhibition of immunosuppressor cells with low doses of an activated analogue of cyclophosphamide and with melphalan has been demonstrated *in vitro*²⁶⁰⁻²⁶³ and *in vivo*.^{263,264} and enhancement of the immune response has been shown *in vivo*.²⁶⁵ For this reason, low doses of cyclophosphamide have been used in conjunction with immunotherapy.^{265,266} Because of its potent immunosuppressive properties, cyclophosphamide has long been used in preparative regimens for allogeneic stem cell transplantation for malignancy²⁶⁷ and more recently for the autologous transplantation of autoimmune disease.^{268,269} The use of high doses of cyclophosphamide without stem cell support has now been reported to produce complete remissions in autoimmune diseases.^{21,270,271}

REFERENCES

- Rhodes R. *The making of the atomic bomb*. New York: Simon & Schuster, 1986.
- Adair CFJ, Bogg HJ. Experimental and clinical studies of the treatment of cancer by dichlorovinylidene (mustard gas). *Ann Surg* 1951;99:190.
- Rhodes C. Nitrogen mustards in treatment of neoplastic disease. *JAMA* 1946;131:6568.
- Goodman LS, Wintrobe MM, Danneberg W, et al. Use of methylbis(benzochloroethyl)hydrochloride for Hodgkin's disease, lymphosarcoma, leukemia. *JAMA* 1946;132:126.
- Jacobson LR, Spurr G, Barron E, et al. Studies of the effect of methylbis(benzochloroethyl)amine hydrochloride on neoplastic diseases and allied disorders of the hematopoietic system. *JAMA* 1946;132:283.
- DeVita VT Jr, Serpick AA, Carbone PP. Combination chemotherapy in the treatment of advanced Hodgkin's disease. *Ann Intern Med* 1973;79:481.
- Millard JT, Rauscher S, Hopkins PB. Mechlorethamine cross links deoxyguanosine residues at 5'-GNG sequences in duplex DNA sequences in duplex DNA fragments. *J Am Chem Soc* 1990;112:2459.
- Brookes P, Lawley PD. The reaction of mono- and difunctional alkylating agents with nucleic acids. *Biochem J* 1961;89:486.
- Dong Q, Borch D, Colvin ME, et al. A structural basis for a phosphoramide mustard-induced DNA interstrand crosslink at 5'-d(GAGC). *Proc Natl Acad Sci USA* 1995;92(26):12170.
- Fisher B, Anderson S, Wickerham DL, et al. Increased intensification and total dose of cyclophosphamide in a doxorubicin-cyclophosphamide regimen for the treatment of primary breast cancer: findings from National Surgical Adjuvant Breast and Bowel Project B-22. *J Clin Oncol* 1997;15(15):1858.
- Falloon G, Torney DC, Carey E, Witte R, Falloon HC. Long-term survival of patients treated with combination chemotherapy for metastatic breast cancer. *Br J Cancer* 1991;78(9):973.
- DeVita VT Jr, Chabner B, Schein P, Hubbard SP, Young RC. Advanced diffuse histiocytic lymphoma, a potentially curable disease. *Lancet* 1975;1:548.
- Chao NJ, Rosenberg SA, Hornig SJ. CEPP(B): an effective and well-tolerated regimen in poor-risk, aggressive non-Hodgkin's lymphoma. *Blood* 1990;76(7):1293.
- Carpenter PB, White L, McCowage G, et al. A dose-intensive, cyclophosphamide-based regimen for the treatment of recurrent/progressive or advanced solid tumors of childhood—a report from the Australia and New Zealand Children's Cancer Study Group. *Cancer* 1997;80(3):489.
- McCowage G, Tien R, McLendon R, et al. Successful treatment of childhood pilocytic astrocytoma metastatic to the leptomeninges with high-dose cyclophosphamide. *Med Pediatr Oncol* 1996;27(1):35.
- Colvin OM. Drug resistance in the treatment of sarcoma. *Semin Oncol* 1997;24(5):580.
- Santos GW, Tutschka PJ, Brookmeyer R, et al. Marrow transplantation for acute nonleukemic leukemia after treatment with busulfan and cyclophosphamide. *N Engl J Med* 1983;309(22):1347.
- Blaize RB, Ramsey NK, Kersey JH, et al. Pretransplant conditioning with busulfan (Myelo-Cy) and cyclophosphamide for nonmalignant diseases. Assessment of engraftment following histocompatible allogeneic bone marrow transplantation. *Transplantation* 1985;39(6):597.
- Antman R, Ayash L, Elias A, et al. High-dose cyclophosphamide, thiopeta, and carboplatin with autologous marrow support in women with measurable advanced breast cancer responding to standard dose therapy: analysis by age. *J Natl Cancer Inst Monogr* 1994;16:91.
- Ferrara F, Copia C, Annunziata M, et al. Complete remission of refractory anemia following a single high dose of cyclophosphamide. *Ann Hematol* 1999;78(2):87.
- Brodsky RA, Peiri M, Smith BD, et al. Immunoblastic high-dose cyclophosphamide without stem-cell rescue for refractory, severe autoimmune disease. *Ann Intern Med* 1998;129(12):1031.
- Colvin HJ. *Pharmacology of cyclophosphamide and metabolites*. *Cancer Treat Rep* 1981;65:89.
- Colvin M, Padgett CA, Fenselau C. A biologically active metabolite of cyclophosphamide. *Cancer Res* 1973;33(4):915.
- Colvin M, Brundrett RB, Kan MN, Jardine I, Fenselau C. Alkylating properties of phosphoramide mustard. *Cancer Res* 1976;36(3):1121.
- Gamick MP, Ludeman SM, Shulman-Roskes EM, et al. Protonation of phosphoramide mustard and other phosphoramides. *J Med Chem* 1993;36(23):3636.
- Hilton J. Deoxyribonucleic acid cross-linking by 4-hydroperoxycyclophosphamide in cyclophosphamide-sensitive and -resistant L1210 cells. *Biochem Pharmacol* 1984;33(12):1867.
- Shulman-Roskes EM, Nee DA, Gamick MP, et al. The partitioning of cyclophosphamide mustard and its activation to form alkylating and PN bond hydrolysis reactions. *J Biol Chem* 1998;273(4):515.
- Hilton J. Role of aldehyde dehydrogenase in cyclophosphamide-resistant L1210 leukemia. *Cancer Res* 1984;44(11):5156.
- Russo JE, Hilton J. Characterization of cytosolic aldehyde dehydrogenase from cyclophosphamide-resistant L1210 cells. *Cancer Res* 1986;46(11):2953.
- Kawar MB, Schlatter E, Russo JE, et al. Direct demonstration of elevated aldehyde dehydrogenase in human hematopoietic progenitor cells. *Blood* 1990;75(10):1947.
- Russo JE, Hilton J, Colvin OM. The role of aldehyde dehydrogenase isozymes in cellular resistance to the alkylating agent cyclophosphamide. *Prog Clin Biol Res* 1989;290:65.
- Nissen-Meyer R, Host H. A comparison between the hematological side effects of cyclophosphamide and nitrogen mustard. *Cancer Chemother Rep* 1980;63:1.
- Mullins GM, Colvin M. Intensive cyclophosphamide (NSC-262771) therapy for solid tumors. *Cancer Chemother Rep* 1975;59(2):411.
- Dong Q, Bullock N, Alloxam F, et al. Reprint analysis of 4-hydroperoxycyclophosphamide induced DNA interstrand cross-linking in the G-Myc gene in 4-hydroperoxycyclophosphamide-sensitive and -resistant medulloblastoma cell lines. *Cancer Chemother Pharmacol* 1996;37(3):242.
- Gamick MP, Mills KK, Colvin M. Noninvasive detection of elevated glutathione levels in MCF-7 cells resistant to 4-hydroperoxycyclophosphamide. *Cancer Res* 1995;55(10):2012.
- Rowley SD, Jones RJ, Piantadosi S, et al. Efficacy of ex vivo purging for autologous bone marrow transplantation in the treatment of acute nonlymphoblastic leukemia. *Blood* 1989;74(1):561.
- Baile JE, Fell VL, Fjellum CR, Thacker EJ. Metabolism of cyclophosphamide by sheep. *J Pharmacol Ther* 1972;20(2):385.
- Jardine I, Fenselau C, Appleg M, et al. Quantitation by gas chromatography-chemical ionization mass spectrometry of cyclophosphamide, phosphoramide mustard, and noninvasive metabolites in the plasma and urine of patients receiving cyclophosphamide therapy. *Cancer Res* 1978;38(2):498.
- Chen TL, Kennedy MJ, Anderson LW, et al. Nonlinear pharmacokinetics of cyclophosphamide and 4-hydroperoxycyclophosphamide/aldophosphamide in patients with metastatic breast cancer receiving high-dose chemotherapy followed by autologous bone marrow transplantation. *Drug Metab Dispos* 1997;25(5):544.
- Ren S, Kallhorn TJ, McDonald GB, et al. Pharmacokinetics of cyclophosphamide and its metabolites in bone marrow transplantation patients. *Clin Pharmacol Ther* 1998;64(3):393.
- Phillips PG, Selevang SB, Cronin AE, PM V. Cyclophosphamide and urinary bladder toxicity. *Cancer Res* 1991;51:1577.
- Forni AM, Koss LG, Geller W. Cytological study of the effect of cyclophosphamide on the epithelium of the urinary bladder in man. *Cancer* 1964;17:1548.
- Cox PJ. Cyclophosphamide cystitis—identification of acrolein as the causative agent. *Br J Clin Pharmacol* 1979;28(1):2045.
- DeFronzo RA, Brindley H, Colvin M, Davis PJ. Water intoxication in man after high-dose therapy. Time course and relation to drug activation. *Ann Intern Med* 1978;76(6):875.
- Harlow PJ, DeClerck VA, Shore NA, et al. Direct cause of inapparent ADH resistance induced by cyclophosphamide therapy. *Cancer* 1979;44(3):896.
- Green TR, Mirkin RL. Prevention of cyclophosphamide-induced antitumor activity by furosemide infusion. *Cancer Therapy* 1981;29(5):634.
- DeFronzo RA, Brindley H, Colvin M, et al. Pathology of high dose intermittent cyclophosphamide therapy. *Hum Pathol* 1975;6(6):693.
- Colvin M. The comparative pharmacology of cyclophosphamide and ifosfamide. *J Clin Oncol* 1982;9(4)(Suppl 1):2.
- Pratt CB, Green AA, Horowitz ME, et al. Central nervous system toxicity following treatment of pediatric patients with ifosfamide/mesna. *J Clin Oncol* 1986;4(8):1253.
- Pratt CB, Meyer WT, Jenkins JJ, et al. Ifosfamide, Fanconi's syndrome, and rickets. *Clin Oncol* 1991(3):1495.
- Boddy AS, Yule SM, Wyllie R, et al. Intrauterine variation in children of ifosfamide: pharmacokinetics and metabolism during repeated administration. *Cancer Chemother Pharmacol* 1996;38(2):147.

58. Boddy AV, Proctor M, Simmonds D, Lind MJ, Idle JR. Pharmacokinetics, metabolism and clinical effect of ifosfamide in breast cancer patients. *Eur J Cancer* 1995;1:69.
59. Ottner J, Wasserman LR, Martel GA, et al. The use of low-dose prednisone and melphalan in the treatment of poor-risk patients with multiple myeloma. *Med Pediatr Oncol* 1975;(12):207.
60. Vose DM, Crowley J, Catchesourian R, et al. High-dose melphalan with autotransplantation for refractory multiple myeloma: results of a southwest oncology group phase II trial. *J Clin Oncol* 1996;17(7):2173.
61. North A, Looi U, Sattray M, Goehli J, Hohenberger W. Pharmacokinetics of melphalan in isolated limb perfusion. *Cancer Chemother Pharmacol* 1999;43(1):35.
62. Goldberger Z, Lee M, Lam HY, Begleiter A. Evidence for carrier-mediated transport of melphalan by L5178Y lymphoblasts in vitro. *Cancer Res* 1977;37(3):755.
63. Begleiter A, Lam HY, Gower C, Frosch E, Goldberger Z. Evidence for active transport of melphalan by two amino acid carriers in L5178Y lymphoblasts in vitro. *Cancer Res* 1979;39(1):353.
64. Vistica DT, Rabon A, Rabinowitz M. Amino acid co-transporter protection against melphalan precursors of amino acids which reduce melphalan toxicity to murine bone marrow precursor cells (CFU-G) and murine L1210 leukemia cells. *Res Commun Chem Pathol Pharmacol* 1979;23(1):171.
65. Groothuis DR, Lippig BE, Fekete I, et al. The effect of an amino acid-lowering diet on the rate of melphalan entry into brain and xenotransplanted glioma. *Cancer Res* 1982;42(20):5690.
66. Falciani SE, Feneau-Gallat M, Krenzel KG, et al. Quantitation by gas chromatography-chemical ionization-mass spectrometry of phenylalanine mustard in plasma of patients. *Cancer Res* 1980;40(7):2268.
67. Herth RK, Ludlum TM, Rubin JC, Knight WA 3rd. Pharmacokinetics of high dose melphalan. *Invest New Drug* 1983;(4):331.
68. Pignatelli B, Martel P, Fabrizio M, et al. Pharmacokinetics of high-dose intravenous melphalan in patients undergoing peripheral blood hematopoietic progenitor-cell transplantation. *Anticancer Res* 1997;17(1B):605.
69. Haas T, Rai RK. Management of chronic lymphocytic leukemia. *Hematol Oncol Clin North Am* 1995;9(2):431.
70. Portlock SC, Fischer DS, Cadman E, et al. High-dose pulse chlorambucil in advanced, low-grade non-Hodgkin's lymphoma. *Cancer Treat Rep* 1987;71(1):1029.
71. Brunen AJW, Reichert LJM, Koene RAP, Wetzels JM. Oral cyclophosphamide versus chlorambucil in the treatment of patients with membranous nephropathy and renal insufficiency. *QM* 1998;91(5):580.
72. Albert DS, Chang SC, Hagg LS, Larson RJ, Evans TL. Comparative pharmacokinetics of chlorambucil and melphalan in man. *Recent Results Cancer Res* 1980;74:124.
73. Przepiorka D, Khouri I, Thall P, et al. Thiopeta, busulfan and cyclophosphamide as a preparative regimen for allogeneic transplantation for advanced chronic myelogenous leukemia. *Res Mater Med Transplant* 1999;23(10):577.
74. Andrichs GJ, Subboshoff LF, Pysagayakaya TI, et al. Direct observation of the alkylation products of deoxyguanosine and DNA by fast atom bombardment mass spectrometry. *Biol Mass Spectrom* 1991;20(11):665.
75. Cohen NA, Egorin MJ, Snyder SW, et al. Interaction of N,N'-N-trichloroethylphosphoramide and N,N'-N-trichloroethylphosphoramide with cellular DNA. *Cancer Res* 1991;51(16):4390.
76. Chang TK, Chen G, Waxman DJ. Modulation of thiopeta antitumor activity in vivo by alteration of liver cytochrome P450-catalyzed drug metabolism. *J Pharmacol Exp Ther* 1978;274(1):270.
77. Kennedy MJ, Armstrong DK, Huelknap AM, et al. Phase I and pharmacologic study of the alkylating agent modulator noraceton in combination with high-dose chemotherapy for the treatment of metastatic breast cancer. *J Clin Oncol* 1995;13(8):1136.
78. Haestlin AM, Petrow WP, Ross M, et al. Phase I/II study of high dose cyclophosphamide, cladribine, and thiopeta followed by autologous bone marrow and granulocyte colony stimulating factor-peripheral blood progenitor cells in patients with advanced malignancies. *Cancer Chemother Pharmacol* 1996;37(6):561.
79. Lysa AF, Luedke S, Einhorn L, Luedke DW, Rusey M. Vincristine and mitomycin C in metastatic breast cancer: A Southeastern Cancer Study Group Trial. *Oncology* 1989;46(6):367.
80. Hong RW, Shen TS, Ko JC, et al. Induction with mitomycin C, doxorubicin, cisplatin and maintenance with weekly 5-fluorouracil, leucovorin for treatment of metastatic nasopharyngeal carcinoma: a phase II study. *Br J Cancer* 1999;80(12):1952.
81. Arbusck SG, Sil Y, Douglas HO Jr, et al. A phase II trial of 5-fluorouracil, doxorubicin, mitomycin C, and leucovorin in advanced gastric carcinoma. *Cancer* 1990;65(11):2442.
82. Borczyk-Borczyk H, Lipman R, Chowdhury D, Tomasz M. Duplex oligodeoxynucleotide cross-linked by mitomycin C at a single site: synthesis, properties, and crosslink reversibility. *Biochemistry* 1990;29(12):2992.
83. Tomasz M, Lipman R, Chowdhury D, et al. Isolation and structure of a covalent cross-link adduct between mitomycin C and DNA. *Science* 1987;235(4793):1204.
84. Rink SM, Lipman R, Alley SC, Hopkins FH, Tomasz M. Binding of DNA by the mitomycin C-Gedexed. CpG intrastatic cross-link. *Chem Res Toxicol* 1996;9(2):382.
85. den Hartog J, Meijer JG, van Oort RP, Fliedert HM. Pharmacokinetics of mitomycin C in humans. *Cancer Res* 1985;45(10):5017.
86. Inatani E, Tamas J. Alkylation by 1,2,5,6-dianhydro-2-thiothymine of deoxyribonucleic acid and guanosine. *Biochem J* 1980;185(3):659.
87. Haas CD, Baker L, Thigpen T. Phase II evaluation of dianhydrothymine in lung cancer: A Southwest Oncology Group Study. *Cancer Ther Res* 1981;36(1):115.
88. Edmonson JH, Frysak S, Lescarier L, Kozik R, Wagner RT. Phase II evaluation of dianhydrothymine in advanced head and neck carcinoma. *Cancer Ther Res* 1979;43(1):3081.
89. Levin VA, Edwards MS, Gutin FH, et al. Phase II evaluation of dianhydrothymine in the treatment of recurrent medulloblastoma, ependymoma, and malignant astrocytoma. *J Neurosurg* 1984;61(6):1068.
90. Nguyen HN, Nordqvist SR. Chemotherapy of advanced and recurrent cervical carcinoma. *Swiss Surg Oncol* 1999;1(3):247.
91. Horvath IF, Cserny J, Kerpel-Fronius S, et al. Pharmacokinetics and metabolism of dianhydrothymine DAG in patients: a comparison with the human disposition of dibromodulcitol DBD. *Eur J Cancer Clin Oncol* 1986;22(2):165.
92. Kelley SL, Peters WF, Andersen J, et al. Pharmacokinetics of dibromodulcitol in humans: a phase I study. *J Clin Oncol* 1986;4(5):753.
93. Hartley JA, Rosenthal MD, Soodam R, et al. An agarose gel method for the determination of DNA interstrand cross-linking applicable to the measurement of the rate of total and "second arm" cross-link reactions. *Anal Biochem* 1991;195(1):131.
94. Streeter RT, Cotter RJ, Colvin ME, Hillon J, Colvin OM. Molecular pharmacology of heparin, NSC 329680: identification of alkylated nucleotides, alkylation site, and site of DNA cross-linking. *Cancer Res* 1995;55(7):1491.
95. Haddad A, Timm GM. Myeloma in chronic myeloid leukemia-chemical constitution and biological action. *Lancet* 1953;2(9):207.
96. Hehlmann R, Heimpel H, Hasford J, et al. Randomized comparison of busulfan and hydroxyurea in chronic myelogenous leukemia: prolongation of survival by hydroxyurea. The German CMG Study Group. *Blood* 1995;82(2):398.
97. Ohnishi K, Tomonaga M, Kamada N, et al. A long term follow-up of a randomized trial comparing interferon-alpha with busulfan for chronic myelogenous leukemia. The Koujicho Leukemia Study Group. *Leuk Res* 1998;22(3):779.
98. Nevill TJ, Barnett MK, Klingemann HG, et al. Regimen-related toxicity of a busulfan-cyclophosphamide conditioning regimen in 70 patients undergoing allogeneic bone marrow transplantation. *J Clin Oncol* 1991;9(7):1224.
99. Santos GW. Busulfan and cyclophosphamide versus cyclophosphamide and total body irradiation for marrow transplantation in chronic myelogenous leukemia—a review. *Leuk Lymph* 1993;1:201.
100. Chao NJ, Stein AS, Long GD, et al. Busulfan/etoposide—initial experience with a new preparative regimen for autologous bone marrow transplantation in patients with acute nonlymphoblastic leukemia. *Blood* 1995;81(2):319.
101. Elion LB. Hematologic effects of the alkylating agents. *Ann NY Acad Sci* 1958;68:826.
102. Haddad JA, Jones RJ, Brundrett RB, et al. Pharmacokinetics of busulfan: correlation with veno-occlusive disease in patients undergoing bone marrow transplantation. *Cancer Chemother Pharmacol* 1989;25(1):55.
103. Jones RJ, Lee KS, Beschornier WE, et al. Venocutaneous disease of the liver following bone marrow transplantation. *Transplantation* 1987;44(6):778.
104. Leiter J, Schneiderman MA. Screening data from the Cancer Chemotherapy National Service Center Screening Laboratories. *Cancer Res* 1980;40(1):31.
105. Johnston TP, McCabe GS, Montgomery JA. The synthesis of antineoplastic agents: XXXII. Nitrofurans. *J Med Chem* 1963;6:669.
106. Montgomery JA. Chemistry and structure-activity studies of the nitrofurans. *Cancer Treat Rep* 1976;60(6):651.
107. Schofield FM. Nitrofurans: a review of experimental antitumor activity. *Cancer Treat Rep* 1976;60(6):665.
108. Schepartz SA. Early history and development of the nitrofurans. *Cancer Treat Rep* 1976;60(6):647.
109. Kohn KW. Interstrand cross-linking of DNA by 1,3-bis(2-chloroethyl)-1-nitrosourea and other 1-(2-haloethyl)-1-nitrosourea. *Cancer Res* 1979;39(7):1450.
110. Colvin M, Brundrett RB, Cowens W, Jardine I, Ludlum DB. A chemical basis for the antitumor activity of chloroethylnitrosoureas. *Biochem Pharmacol* 1976;25(6):959.
111. Tong WP, Kirk MK, Ludlum DB. Mechanism of action of the nitrosoureas. V. Formation of O⁶-(2-fluoroethyl)guanine and its probable role in the cross-linking of deoxyribonucleic acid. *Biochem Pharmacol* 1983;32(15):2011.
112. Fischhaber PL, Gall AS, Duncan JA, Hopkins PB. Direct demonstration in synthetic oligonucleotides that N⁷-bis(2-chloroethyl)-nitrosourea cross-links N³ of deoxyguosine to N³ of deoxythymine on opposite strands of duplex DNA. *Cancer Res* 1991;50(17):4583.
113. Walker MD, Alexander E Jr, Hunt WE, et al. Evaluation of BCNU and/or carboxymethyl in the treatment of anaplastic glioma. A cooperative clinical trial. *J Neurosurg* 1978;49(3):333.
114. Bladt J, Rozman C, Monsterrat E, et al. Treatment of alkylation resistant multiple myeloma with vincristine, BCNU, doxorubicin and prednisone (VABP). *Eur J Cancer Clin Oncol* 1988;22(10):1193.
115. Elder JR, Asman R, Peters W, et al. High-dose combination alkylating agent chemotherapy with autologous bone marrow support for metastatic breast cancer. *J Clin Oncol* 1986;4(11):1592.
116. Garfield J, Dwyer AD, Weller RO. Postoperative intracavitary chemotherapy of malignant supratentorial astrocytomas using BCNU. *Clin Oncol* 1975;1(3):213.
117. Bren H, Piantadosi S, Burger PC, et al. Placebo-controlled trial of safety and efficacy of intraperitoneal controlled delivery by intracavitary polymers of chemotherapy for recurrent glioma. *Lancet* 1995;345(8986):1008.
118. Paleologos NA, Macdonald DR, Vick NA, Cairncross JG. Neoadjuvant procarbazine, CCNU, and vincristine for anaplastic and aggressive oligodendroglioma. *Neurology* 1995;55(3):1161.
119. Fowler M, Goss C, Curran WJ, et al. Procarbazine, lomustine, and vincristine (PCV) chemotherapy for anaplastic astrocytoma: a retrospective review of Radiation Therapy Oncology Group protocols comparing survival with carmustine or PCV adjuvant chemotherapy. *J Clin Oncol* 1999;17(11):3389.
120. Clark JF, Barcaccia P, Nava HR, Goodwin PS, Douglas HS. Adjuvant 5-FU and MCCNU improves survival following curative gastrectomy for adenocarcinoma. *Am Surg* 1990;56(7):527.
121. Paccapelo A, Piana G, Rycklicki F, et al. Treatment of malignant gliomas: a new approach. *Tumori* 1998;84(5):593.
122. Arita N, Ushio Y, Hayakawa T, et al. Intrathecal ACNU—a new therapeutic approach against malignant leptomeningeal metastases. *J Neurooncol* 1988;6(3):221.
123. Alberts DS, Durie BG, Salmon SE. Doxorubicin/B.G.N.U. chemotherapy for multiple myeloma in relapse. *Lancet* 1976;1(7966):926.

118. Levin VA, Hoffman W, Weisman RJ. Pharmacokinetics of BCNU in man: a preliminary study of 30 patients. *Cancer Treat Res* 1956;62(9):1305.
119. Meisenberg BR, Rose M, Vredenburg JJ, et al. Randomized trial of high-dose chemotherapy with autologous bone marrow support as adjuvant therapy for high-risk, multi-node-positive malignant melanoma. *J Natl Cancer Inst* 1993;85(11):1080.
120. Henner WD, Peters WF, Elder JP, et al. Pharmacokinetics and immediate effects of high-dose carmustine in man. *Cancer Treat Res* 1986;70:1877.
121. Levin VA, Stenzel J, Byrd A, Finn A, Weisman RJ. The effect of phenobarbital pretreatment on the antitumor activity of 1,3-bis(2-chloroethyl)-1-nitrosourea (BCNU), 1-(2-chloroethyl)-3-cydoheptyl-1-nitrosourea (CCNU) and 1-(2-chloroethyl)-5-(2,6-dioxo-3-piperidinyl)-1-nitrosourea (PCNU), and on the plasma pharmacokinetics and biotransformation of BCNU. *J Pharmacol Exp Ther* 1979;208(1):1.
122. Lee FY, Worlman P, Roberts TJ, Bleehen NM. Clinical pharmacokinetics of oral CCNU (lomequinox). *Cancer Chemother Pharmacol* 1986;14(2):135.
123. DeVita VT, Carbone PP, Owens AH Jr, et al. Clinical trials with 1,3-bis(2-chloroethyl)-1-nitrosourea, NSC-409962. *Cancer Res* 1965;25:1876.
124. Toth B. Synthetic and naturally occurring hydrazines as possible cancer causative agents. *Cancer Res* 1975;35(12):3693.
125. Silverstein R, Bhatia P, Svoboda IJ. Effect of hydropyridine sulfate on glucose-regulating enzymes in the normal and cancerous rat. *Toxicopharmacology* 1989;17(1):37.
126. Gold J. Use of hydropyridine sulfate in terminal and preterminal cancer patients: results of investigational new drug (IND) study in 84 evaluable patients. *Oncology* 1975;32(1):1.
127. Herndon JE, Fleishman S, Kost MY, Green MR. A longitudinal study of quality of life in advanced non-small cell lung cancer—Cancer and Leukemia Group B 8931. *Control Clin Trials* 1997;18(4):236.
128. Ramnath JM, Fietz KJ. The effect of hydropyridine sulfate on prostate cancer growth. *Oncol Rep* 1998;5(4):919.
129. Spies SK, Synman HW. Procarbazine (Nalutane) in the treatment of Hodgkin's disease and other lymphomas. *S Afr Med J* 1966;40(4):1061.
130. Glick JH, Young ML, Harrington D, et al. MOFP/ABV hybrid chemotherapy for advanced Hodgkin's disease significantly improves failure and overall survival: the 8-year results of the intergroup trial. *J Clin Oncol* 1998;16(14):4139.
131. Brundage AA, Ermani M, Turzai S, et al. Procarbazine and high-dose tamoxifen as a second-line regimen in recurrent high-grade glioma: phase II study. *J Clin Oncol* 1999;17(2):645.
132. Fink D, Aebi S, Howell SB. The role of DNA mismatch repair in drug resistance. *Clin Cancer Res* 1998;4(1):1.
133. Friedman HS, Johnson SF, Dong Q, et al. Methylator resistance mediated by mismatch repair deficiency in a glioblastoma xenograft. *Cancer Res* 1997;57(14):2933.
134. Bianchini F, Weidarpas E, Kyriopoulos S, et al. Detection of DNA methylation adducts in Hodgkin's disease patients treated with procarbazine. *Biomarkers* 1996;1(4):226.
135. Erikson JM, Tweedie DJ, Ducrocq JM, Prough RA. Cytotoxicity and DNA damage caused by the azoxy metabolites of procarbazine in L1210 mouse cells. *Cancer Res* 1989;49(1):127.
136. Swaffey DS, Horstman MG, Jew JF, et al. Methylglutathione-procarbazine, the active metabolite responsible for the anticancer activity of procarbazine against L1210 leukemia. *Cancer Res* 1989;49(24):242.
137. Masie MJ, Holland JC. Diagnosis and treatment of depression in the cancer patient. *J Clin Psychiatry* 1986;47(3):25.
138. Pfefferbaum B, Pack R, van Eys J. Monoamine oxidase inhibitor toxicity. *J Am Acad Child Adolesc Psychiatry* 1989;28(6):954.
139. Ferina P, Beninati E, Reggiani R, et al. Metabolism of the anticancer agent 1-(4-acephylphenyl)-3,5-dimethylpyrazine. *Biomol Mass Spectrom* 1983;10(8):485.
140. Skibba LJ, Best DD, Ramirez G, Bryan GT. N-demethylation of the antineoplastic agent(5)-(3,5-dimethyl-1-oxazolo-5-yl)-4-carboxamide by rat and man. *Cancer Res* 1970;30(1):47.
141. Vaughan K, Tang Y, Llanos G, et al. Studies of the mode of action of antitumor triazines and triazines: 6-1-Aryl-3-(hydroxymethyl)-5-methylthio-1,2,4-triazines: synthesis, chemistry, and antitumor properties. *J Med Chem* 1984;27(3):357.
142. DeVita VT, Mauch PM, Harris NL. Hodgkin's disease. In: DeVita VT, Hellman S, Rosenberg S, eds. *Cancer: principles of practice and oncology*. Philadelphia: Lippincott-Raven Publishers, 1997:2608.
143. Falkson G, Ibrahim I, Kirkwood JM, et al. Phase III trial of dacarbazine versus dacarbazine with interferon alpha-2b versus dacarbazine with tamoxifen versus dacarbazine with interferon alpha-2b and tamoxifen in patients with metastatic malignant melanoma: an Eastern Cooperative Oncology Group study. *J Clin Oncol* 1986;15(1):1745.
144. Lowe PR, Sansom CE, Schwabe CH, Stevens MF, Clark AS. Antitumor immunotaxanes: 25. Crystal structure of 8-carboxamido-5-methylimidazo[5,1-d]-1,2,3,5-tetrazin-4(3H)-one (temozolomide) and structural comparisons with the related drugs mitozolomide and DTIC. *J Med Chem* 1992;35(18):3377.
145. Deney RJ, Wheelhouse RT, Stevens MF, Tang LL, Slack JA. NMR and molecular modeling investigation of the mechanism of activation of the antitumor drug temozolomide and its interaction with DNA. *Biochemistry* 1994;33(31):9045.
146. Nicholson HS, Krailo M, Ames MM, et al. Phase I study of temozolomide in children and adolescents with recurrent solid tumors—a report from the Children's Cancer Group. *J Clin Oncol* 1998;16(9):3037.
147. Hammond LA, Eckardt JR, Baker SD, et al. Phase I and pharmacokinetic study of temozolomide on a daily-for-5-days schedule in patients with advanced solid malignancies. *J Clin Oncol* 1997;15(26):2604.
148. Paulsen F, Hoffmann W, Becker G, et al. Chemotherapy in the treatment of recurrent glioblastoma multiforme: flutamide versus temozolomide. *J Cancer Res Clin Oncol* 1999;125(7):411.
149. Newlands ES, O'Reilly SM, Glaser MG, et al. The Charing Cross Hospital experience with temozolomide in patients with gliomas. *Eur J Cancer* 1996;32:2236.
150. Wolf RJ, Judson I, Lee SM, et al. Temozolomide in adult patients with advanced soft tissue

- sarcoma: a phase II study of the EORTC Soft Tissue and Bone Sarcoma Group. *Eur J Cancer* 1999;35(3):410.
151. Moore MJ, Feld R, Hedley D, Oza A, Siu LL. A Phase II study of temozolomide in advanced untreated pancreatic cancer. *Int J Cancer* 1998;16(1):177.
152. Beiruthaupt H, Dummann A, Aigner K. Pharmacokinetics of dacarbazine (DTIC) and its metabolite 5-aminoimidazole-4-carboxamide (AIC) following different dose schedules. *Cancer Chemother Pharmacol* 1981;7(2):103.
153. Adkins DR, Irwin R, Kuhn J, et al. A phase I clinical and pharmacological profile of dacarbazine with autologous bone marrow transplantation in patients with solid tumors. *Hum Nau Drugs* 1993;11(2):160.
154. Reid JM, Stevens DC, Rubin J, Ames MM. Pharmacokinetics of 3-methyl-(triazene-1-yl)-imidazole-4-carboxamide. *Clin Cancer Res* 1997;3(12):2399.
155. Baker SD, Wirth M, Sawicki H, et al. Absorption, metabolism, and excretion of 1-(5-oxo-5H-tetrazol-5-yl)-3-methyl-1H-imidazole-4-carboxamide following oral administration to patients with advanced cancer. *Clin Cancer Res* 1995;1(2):309.
156. Friedman HS, Colvin OM, Kaufmann SH, et al. Cyclophosphamide resistance in medulloblastoma. *Cancer Res* 1992;52(19):5373.
157. Sundtshoek K, Petro BJ, Vistica DT. Reduction in glutathione content of L-PAM resistant L1210 cells confers drug sensitivity. *Biochem Pharmacol* 1982;31(1):121.
158. Butler AL, Clapper ML, Tew KD. Glutathione S-transferases in nitrogen mustard-resistant and -sensitive cell lines. *Mol Pharmacol* 1987;31(6):575.
159. Puchalski RB, Fahl WE. Expression of recombinant glutathione S-transferase γ_1 , γ_2 , or γ_3 confers resistance to alkylating agents. *Proc Natl Acad Sci U S A* 1990;87(7):2443.
160. Townsend M, Peicki WR, Haynes RL, et al. Characteristic functions of glutathione S-transferases in cell lines induced to express specific isoenzymes by stable transfection. *Clin Biol Invest* 1998;112:389.
161. Dulik DM, Colvin OM, Feneau C. Characterization of glutathione conjugates of chlorambucil by fast atom bombardment and thermogravimetric liquid chromatography/mass spectrometry. *Biomol Environ Mass Spectrom* 1990;19(4):248.
162. Dulik DM, Feneau C, Hilton J. Characterization of melphalan-glutathione adducts with glutathione conjugated by glutathione transferase. *Biochem Pharmacol* 1986;35(19):2405.
163. Yuan ZM, Feneau C, Dulik DM, et al. Laser desorption electron impact application to a study of the mechanism of conjugation of glutathione and cyclophosphamide. *Anal Chem* 1990;62(8):868.
164. Bolton MG, Colvin OM, Hilton J. Specificity of isoenzymes of murine hepatic glutathione S-transferase for the conjugation of glutathione with L-phenylalanine mustard. *Cancer Res* 1991;51(20):2840.
165. Giacino PJ, Tew KD, LaCreta PF. The spontaneous and glutathione S-transferase-mediated reaction of chlorambucil with glutathione. *Cancer Comm* 1990;2(6):279.
166. Divren HA, van Ommer B, van Bladeren PJ. Involvement of human glutathione S-transferase isoenzymes in the conjugation of cyclophosphamide metabolites with glutathione. *Cancer Res* 1994;54(23):6201.
167. Pallares SL, Lick CA, Dulik DM, Feneau C. Glutathione conjugates. Immobilized enzyme synthesis and characterization by fast atom bombardment mass spectrometry. *Drug Metab Dispos* 1986;14(3):313.
168. Horton JN, Roy G, Piper JT, et al. Characterization of a chlorambucil-resistant human ovarian carcinoma cell line overexpressing glutathione S-transferase μ_1 . *Biochem Pharmacol* 1990;58(4):603.
169. Anderson ME. Glutathione: an overview of biosynthesis and modulation. *Clin Biol Invest* 1998;112:1.
170. Smith AC, Liao JT, Page JG, Wientjes MG, Grishaber CK. Pharmacokinetics of buthionine sulfoximine (NCS 326831) and its effect on melphalan-induced toxicity in mice. *Cancer Res* 1989;49(19):5586.
171. Friedman HS, Colvin OM, Jaffe OW, et al. Increased melphalan-induced toxicity in intracranial human medulloblastoma and glioma xenografts following buthionine sulfoximine-mediated glutathione depletion. *J Natl Cancer Inst* 1989;81(7):524.
172. Bailey HH, Rippl G, Tusch KD, et al. Phase I study of continuous-infusion L-SR buthionine sulfoximine with intravenous melphalan. *J Natl Cancer Inst* 1997;89(23):1769.
173. Morgan AS, Gascio PL, Tew KD, Kaurav LN. Isotype specific glutathione S-transferase inhibitors potentiate drug sensitivity in cultured human tumor cell lines. *Cancer Chemother Pharmacol* 1997;41(3):363.
174. Zhang K, Yang FB, Wong KP, Mack P, GSH, GST-related enzymes and GSH pump in relation to sensitivity of human tumor cell lines to chlorambucil and adriamycin. *Int J Oncol* 1999;14(5):861.
175. Gupta V, Jani JP, Jacob S, et al. Activity of melphalan in combination with the glutathione transferase inhibitor sulfinylsulfonamide. *Cancer Chemother Pharmacol* 1995;36(1):15.
176. Keppeler D, Leier I, Jellitschky G, Kooij J. ATP-dependent transport of glutathione S-conjugates by the multidrug resistance protein Mrp1 and its apical isoform Mrp2. *Clin Biol Invest* 1998;112:153.
177. Barouqui K, Leier I, Jellitschky G, et al. Multidrug resistance protein-mediated transport of chlorambucil and melphalan conjugated to glutathione. *Br J Cancer* 1998;77(8):201.
178. Kelley S, Bana A, Tricher BA, et al. Overexpression of metallothionein confers resistance to anticancer drugs. *Semin Oncol* 1984;11(4):1813.
179. Yu X, Wu Z, Feneau C. Covalent conjugation of melphalan by metallothionein and selective alkylation of cysteines. *Biochemistry* 1995;34(10):3377.
180. Wei D, Fabris D, Feneau C. Covalent sequestration of phosphoramide mustard by metallothionein—in a *visu* study. *Drug Metab Dispos* 1999;27(7):766.
181. Sato M, Chertan MG, Imura N, Shimizu H. Modulation of resistance to anticancer drugs by inhibition of metallothionein synthesis. *Cancer Res* 1994;54(20):5255.
182. Pegg AE, Boosalis M, Samson L, et al. Mechanism of inactivation of human O⁶-allylguanine-DNA alkyltransferase by O⁶-benzylguanine. *Biochemistry* 1993;32(45):11998.

185. Pegg AE. Mammalian O⁶-allylguanine-DNA alkyltransferase: regulation and importance in response to alkylating carcinogenic and therapeutic agents. *Cancer Res* 1990;50(19):6119.
186. Erickson LC, Laurent G, Sharkey NA, Kohn KW. DNA cross-linking and monoduct repair in nitrosourea-treated human tumor cells. *Nucleic Acids Res* 1980;8(7):2792.
187. Bodel WJ, Tobolski K, Ladum D. Differences in DNA alkylation products formed in sensitive and resistant human glioma cells treated with N-(2-chloroethyl)-N-nitrosourea. *Cancer Res* 1988;48(16):4489.
188. Dolan ME, Moschel RC, Pegg AE. Depletion of mammalian O⁶-allylguanine-DNA alkyltransferase activity by O⁶-benzylguanine provides a means to evaluate the role of this protein in protection against carcinogenic and therapeutic alkylating agents. *Proc Natl Acad Sci U S A* 1990;87(14):5088.
189. Gerson SL, Berger SL, Varnes ME, Donovan C. Combined depletion of O⁶-allylguanine-DNA alkyltransferase and glutathione to modulate nitrosourea resistance in breast cancer. *Cancer Pharmacol* 1994;48(3):543.
190. Csanac C, Rapp M, Moustouros K, et al. Enhancement by O⁶-benzyl-N-acetylguanine derivatives of chloroethylnitrosourea antitumor action in chloroethylnitrosourea-resistant human malignant melanocytes. *J Pharmacol Exp Ther* 1994;271(3):553.
191. Friedman HS, Kolkmanik MD, Pluda J, et al. Phase I trial of O⁶-benzylguanine for patients undergoing surgery for malignant gliomas. *J Clin Oncol* 1996;14(11):3570.
192. Spiro TP, Gerson SL, Liu LL, et al. O⁶-benzylguanine: a clinical trial establishing the biochemical modulatory dose in tumor tissue for alkyltransferase-directed DNA repair. *Cancer Res* 1999;59(10):2402.
193. Allan JA, Duenkel LL, Koc ON, Liu L, Gerson SL. Retroviral transduction and expression of the human alkyltransferase cDNA provides nitrosourea resistance to hematopoietic cells. *Blood* 1995;85(11):3342.
194. Koc ON, Reese JS, Davis BM, et al. Delta MGMT-Transduced bone marrow infusion increases tolerance to O⁶-benzylguanine and 1,3-bis(2-chloroethyl)-1-nitrosourea and allows intensive therapy of 1,3-bis(2-chloroethyl)-1-nitrosourea-resistant human colon cancer xenografts. *Hum Gene Ther* 1999;10(6):1021.
195. Kohn KW, Steigbigel NH, Spears KL. Cross-linking and repair of DNA in sensitive and resistant strains of *E. coli* treated with nitrogen mustard. *Proc Natl Acad Sci U S A* 1984;81(3):1154.
196. Grathorn AR, Roberts JJ. Mechanism of the cytotoxic action of alkylating agents in mammalian cells and evidence for the removal of alkylated groups from deoxyribonucleic acid. *Nucleic Acids Res* 1962;11(4):130.
197. Sancar A. Mechanisms of DNA excision repair. *Science* 1994;266(5193):1954.
198. Steamer T, Ding R, Smulmon M, Bohr VA. Inhibition of gene-specific repair of alkylated damage in cells depleted of poly(ADP-ribose) polymerase. *Nucleic Acids Res* 1994;22(22):4620.
199. Das SK, Laszlo GC, Pardee A. Comparative analysis of caffeine and 5-aminomethylbenzamide as DNA repair inhibitors in *S. typhimurium* hamster kidney cells. *Mutat Res* 1984;131(2):71.
200. O'Connor PM, Ferris DK, White GA, et al. Relationships between o62 lesions, DNA cross-linking, and cell cycle perturbations induced by nitrogen mustard. *Cell Growth Differ* 1992;3(1):43.
201. O'Connor PM, Ferris DK, Hoffman L, et al. Role of the cdc25C phosphatase in G2 arrest induced by nitrogen mustard. *Proc Natl Acad Sci U S A* 1994;91(20):9480.
202. Selby CF, Sancar A. Molecular mechanisms of DNA repair inhibition by caffeine. *Proc Natl Acad Sci U S A* 1980;77(9):3022.
203. Walton MI, Whynop D, O'Connor PM, et al. Constitutive expression of human hC3 modulates nitrogen mustard and camptothecin induced apoptosis. *Cancer Res* 1995;55(8):1853.
204. Dong Q, Johnson SF, Colvin OM, et al. Multiple DNA repair mechanisms and alkylator resistance in the human melanoblastoma cell line D-283 Mel (4-HCR). *Cancer Chemother Pharmacol* 1999;45(1):73.
205. Kobayashi H, Man S, Graham CH, et al. Acquired multidrug-resistance to alkylating agents in cancer. *Proc Natl Acad Sci U S A* 1993;90(8):3494.
206. St Croix B, Man S, Kerbel RS. Reversal of intrinsic and acquired forms of drug resistance by hyaluronidase treatment of solid tumors. *Cancer Lett* 1988;151(1):35.
207. Kerbel RS, Kobayashi H, Graham CH. Intrinsic or acquired drug resistance and metastasis are they linked phenomena? *Cell Growth Differ* 1994;5(4):37.
208. Mullins GM, Anderson FN, Santos GW. High dose cyclophosphamide therapy in solid tumors. Therapeutic, toxic, and immunosuppressive effects. *Cancer* 1990;56(6):1950.
209. Fried W, Kedo A, Barone J. Effects of cyclophosphamide and of busulfan on spleen colony formation and on hematopoietic stroma. *Cancer Res* 1977;37(4):1205.
210. Borison HL, Brand ED, Orland RK. Emetic action of nitrogen mustard in dogs and cats. *Am J Physiol* 1968;216:110.
211. Fetting JH, McCurdy LE, Borison HL, Colvin M. Vomiting induced by cyclophosphamide and phosphoramide mustard in rats. *Cancer Treat Res* 1982;56(8):1625.
212. Spitzer TR, Grunberg SM, Dicato MA. Antiemetic strategies for high-dose chemoradiotherapy-induced nausea and vomiting. *Support Care Cancer* 1998;6(3):233.
213. Perez EA. 5-HT₃ antiemetic therapy for patients with breast cancer. *Breast Cancer Res Treat* 1999;57(2):207.
214. Beaudet F, Coiffier B, Desablens B. Granisetron plus or minus aprepitant for emesis prevention in chemotherapy of lymphoma: a randomized multicenter trial. *Leuk Lymph* 1999;34(4):541.
215. Anuman K, Eder JP, Elias A, et al. High-dose thiotepa alone and in combination regimens with bone marrow support. *Semin Oncol* 1990;17(Suppl 3):35.
216. Spitz S. The histological effects of nitrogen mustard on human tumors and tissues. *Cancer* 1948;1:383.
217. Miller DG. Alkylating agents and human spermatogenesis. *JAMA* 1971;212(12):1662.
218. Sherins RJ, DeVita VT Jr. Effect of drug treatment for lymphoma on male reproductive capacity: Studies in men in remission after therapy. *Ann Intern Med* 1973;79(2):216.
219. Hiale DA, Heller HJ, Hsu SH, Schacter BG. Return of fertility in a patient with cyclophosphamide-induced azoospermia. *Johns Hopkins Med J* 1976;139(1):80.
220. Hinkes E, Pothof D. Reversible drug-induced sterility in a patient with acute leukemia. *JAMA* 1960;223(13):1490.
221. Galton DAG, Till M, Withshaw E, Boualfar B. 1,4-dimethanesulfonyloxypentane. Myelocrit: summary of clinical results. *Ann NY Acad Sci* 1958;68:167.
222. Rose DJ, Davis TE. Ovarian function in patients receiving adjuvant chemotherapy for breast cancer. *Lancet* 1973;1(8028):1174.
223. Koyama H, Wada T, Nishizawa Y, Inagawa T, Aoki Y. Cyclophosphamide-induced ovarian failure and its therapeutic significance in patients with breast cancer. *Cancer* 1985;55(4):1405.
224. Oliner H, Schwartz R, Rubio JP. Interstitial pulmonary fibrosis following busulfan therapy. *Am Med* 1961;31:134.
225. Gidding RW, Chabers TM. Pulmonary fibrosis following therapy with melphalan for multiple myeloma. *J Clin Pathol* 1972;25(8):668.
226. Cole SR, Myers TJ, Klatky AU. Pulmonary disease with chlorambucil therapy. *Cancer* 1978;41(2):467.
227. Mark GJ, Lehman-Zadeh A, Rapoport BD. Cyclophosphamide pneumonitis. *Thorax* 1978;33(1):89.
228. Patel AR, Shuc C, Rhee HL, Sassoon H, Rao KP. Cyclophosphamide therapy and interstitial pulmonary fibrosis. *Cancer* 1976;38(4):1542.
229. Orwoll ES, Kieselring JP, Patterson JR. Interstitial pneumonitis from mitomycin. *Ann Intern Med* 1978;89(3):392.
230. Bailey CG, Marsden HB, Jones PH. Fatal pulmonary fibrosis following 1,3-bis(2-chloroethyl)-1-nitrosourea (BCNU) therapy. *Cancer Res* 1978;42(1):74.
231. Hopley PE, Jenkins DM, Greenberg SD. Pulmonary toxicity in long-term administration of BCNU. *Cancer Treat Res* 1976;60(1):1501.
232. Litam JP, Dai DH, Spitzer G, et al. Early pulmonary toxicity after administration of high-dose BCNU. *Cancer Treat Res* 1981;65(1):39.
233. Wilczynski SW, Erasmus JJ, Petros WP, Vredenburg JP, Folz RJ. Delayed pulmonary toxicity syndrome following high-dose chemotherapy and bone marrow transplantation for breast cancer. *Am J Hosp Med* 1998;15(7):568.
234. Cobin M, Cowen JW, Brundrett BR, Kramer BS, Ladum DB. Decomposition of BCNU (1,3-bis(2-chloroethyl)-1-nitrosourea) in aqueous solution. *Biochem Biophys Res Commun* 1974;60(2):515.
235. Vijayan VR, Sankaran K. Relationship between lung inflammation, changes in lung function and severity of exposure in victims of the Bhopal tragedy. *Env Res* 1994;62(1):107.
236. Bierman RH, Kelly KH, Knudson AG Jr, MacKawa T, Timmis GM. The influence of 1,4-dimethylsulfonyl-1,4-dimethylbutane (CS 2348, dimethylmyleran) in neoplastic disease. *Ann NY Acad Sci* 1984;428:1221.
237. Fei YJ, Lamoureux CH. Allopurinol activity of cyclophosphamide metabolites and related compounds in sheep. *Cancer Res* 1974;34(10):2596.
238. O'Connor PM, Ferris DK, White GA, et al. A comparison of the effects of various nitrogen mustard compounds on embryonic cells. *J Exp Med* 1992;175:108:75.
239. Murphy ML, Del Moro A, Lacon C. The comparative effects of five polycyclic alkylating agents on the rat fetus, with additional notes on the chick embryo. *Ann NY Acad Sci* 1958;68:762.
240. Hales RF. Effects of phosphoramide mustard and acrolein, cytotoxic metabolites of cyclophosphamide, on mouse limb development in vitro. *Toxicity* 1980;40(1):11.
241. Mirkes PE. Cyclophosphamide teratogenesis: a review. *Teratog Carcinog* 1985;5(2):75.
242. Nicholson HO. Cytotoxic drugs in pregnancy: Review of reported cases. *J Obstet Gynaecol Br Comm* 1968;75(3):307.
243. Lergier JR, Jimenez E, Maldonado N, Versy F. Normal pregnancy in multiple myeloma treated with cyclophosphamide. *Cancer* 1974;34(4):1018.
244. Orsini J. Multiple agent chemotherapy including bleomycin in non-Hodgkin's lymphoma during pregnancy. *Cancer* 1977;40(2):2829.
245. Reichman BS, Green KB. Breast cancer in young women: effect of chemotherapy on ovarian function, fertility, and birth defects. *J Natl Cancer Inst* 1994;86:16:25.
246. Aviles A, Diaz-Maqueo JC, Talavera A, Guzman R, Garcia LL. Growth and development of children of mothers treated with chemotherapy during pregnancy: current status of 43 children. *Am J Humatol* 1991;36(4):524.
247. Hochberg MG, Shulman LE. Acute leukemia following cyclophosphamide therapy for Sjögren's syndrome. *Johns Hopkins Med J* 1978;142(6):211.
248. Ruzer F, Grunwald H. Multiple myeloma terminating in acute leukemia. Report of 12 cases and review of the literature. *Am J Med* 1974;57(4):927.
249. Ruzer F, Grunwald H. Hodgkin's disease terminating in acute leukemia. Report of eight cases and review of the literature. *Am J Med* 1975;58(3):339.
250. Einhorn A. Acute leukemia after chemotherapy (melphalan). *Cancer* 1978;41(2):444.
251. Reimer RR, Hoover R, Fraumeni JF Jr, Young RC. Acute leukemia after alkylating-agent therapy of ovarian cancer. *N Engl J Med* 1977;297(4):177.
252. Greene MH, Harris EL, Gershenson DM, et al. Melphalan may be a more potent leukemogen than cyclophosphamide. *Ann Intern Med* 1986;105(3):360.
253. Einhorn B, Elund G, Lambert B. Solid tumours and chromosome aberrations as late side effects of melphalan therapy in ovarian carcinoma. *Acta Oncol* 1988;27(2):215.
254. Tucker MA, Coleman CN, Cox RS, Varghese A, Rosenberg SA. Risk of second cancers after treatment for Hodgkin's disease. *N Engl J Med* 1988;318(2):76.
255. Heikinen L, Cooper HJ. The effect of mustard gas (dichlorodiphosphoride) on antibody formation. *J Hyg Med* 1921;28:279.
256. Makinodan T, Santos GW, Quinn RE. Immunosuppressive drugs. *Pharmacol Rev* 1970;22(1):80.
257. Barratt TM, Southall JE. Controlled trial of cyclophosphamide in steroid-sensitive relapsing nephrotic syndrome of childhood. *Lancet* 1970;2(7671):479.
258. Laros RK, Pomeroy JA. "Refractory" thrombocytopenic purpura treated successfully with cyclophosphamide. *JAMA* 1971;215(3):445.
259. Klenz R. Cyclophosphamide and methoprednisolone sulfate therapy for minimal lesion glomerulonephritis. *Kidney Int* 1995;56(6):2312.

258. Bargman JM. Management of minimal lesion glomerulonephritis: evidence-based recommendations. *Kidney Int* 1999;55(Suppl 70):S3.
259. Viallard JF, Pellegrin JL, Vergnes C, et al. Three cases of acquired von Willebrand disease associated with systemic lupus erythematosus. *Br J Haematol* 1999;105(2):532.
260. Ozer H, Coenen W, Colvin M, Nussbaum-Blumenthal A, Sheedy D. In vitro effects of 4-hydroperoxycyclophosphamide on human immunoregulatory T subset function. I. Selective effects on lymphocyte function in T-B cell collaboration. *J Exp Med* 1982; 155(1):276.
261. Smith JJ, Mihich E, Ozer H. In vitro effects of 4-hydroperoxycyclophosphamide on human immunoregulatory T subset function. *Methods Find Exp Clin Pharmacol* 1987;9(9):355.
262. Mokry MB, Colvin M, Drey S. Cyclophosphamide-mediated enhancement of antitumor immune potential of immunosuppressed spleen cells from mice bearing a large MOPC-315 tumor. *Int J Immunopharmacol* 1985;7(1):111.
263. Drey S, Mokry MB. Cyclophosphamide and melphalan as immunopotentiating agents in cancer therapy. *Med Oncol Tumor Pharmacother* 1989;6(1):77.
264. Berr D, Mastrangelo MJ. Effect of low dose cyclophosphamide on the immune system of cancer patients: depletion of CD4+, 2E14+ suppressor-inducer T cells. *Cancer Res* 1988;48(6):1671.

STEVEN W. JOHNSON
JAMES P. STEVENSON
PETER J. O'DWYER

SECTION 4

Cisplatin and Its Analogues

The platinum drugs represent a unique and important class of antitumor compounds. Alone or in combination with other chemotherapeutic drugs, *cis*-diamminedichloroplatinum (II) (cisplatin) and its analogues have made a significant impact on the treatment of a variety of solid tumors. The realization that platinum complexes exhibit antitumor activity arose somewhat serendipitously in a series of experiments carried out by Rosenberg and colleagues beginning in 1961.¹ These studies involved determining the effect of electromagnetic radiation on the growth of bacteria in a chamber equipped with a set of platinum electrodes. Exposure of the bacteria to an electric field resulted in a profound change in their morphology and, in particular, the appearance of long filaments that were several hundred times longer than that of their untreated counterparts. This effect was not due to the electric field directly, but to the electrolysis products produced from the platinum electrodes. An analysis of these products revealed that the predominant species was ammonium chloroplatinate $[NH_4]_2[PtCl_6]$. This compound was inactive at reproducing the filamentous growth originally observed; however, Rosenberg and colleagues soon discovered that the conversion of this complex to a neutral species by UV light resulted in an active species. Attempts to synthesize the active neutral platinum complex failed. They realized, however, that the neutral compound could exist in two isomeric forms, *cis* or *trans*, and that the latter species is the one they had synthesized. Subsequently, the *cis* isomer was synthesized and shown to be the active compound.

The observation that *cis*-diamminedichloroplatinum (II) and *cis*-diamminetetrachloroplatinum (IV) inhibited bacterial growth led to the testing of four neutral platinum compounds for antineoplastic activity in mice bearing the Sarcoma-180 solid tumor and L1210 leukemia cells.² All four compounds showed significant antitumor activity, with *cis*-diamminedichloroplatinum (II) exhibiting the most efficacy. Further studies in other tumor models confirmed these results and indicated that cisplatin exhibited a broad spectrum of activity. Although early

265. Berr D, Mastrangelo MJ. Active immunotherapy of human melanoma exploiting an immunopotentiating effects of cyclophosphamide. *Cancer Invest* 1988;6(3):357.
266. Mitchell MS, Kempf RA, Hard W, et al. Effectiveness and tolerability of low-dose cyclophosphamide and low-dose intravenous interferon- α in disseminated melanoma. *J Clin Oncol* 1988;6(4):402.
267. Santos GW, Senesbrenner LL, Anderson PN, et al. HLA-identical marrow transplant in aplastic anemia, acute leukemia, and lymphosarcoma employing cyclophosphamide. *Transplant Proc* 1976;8(4):607.
268. Burt RK, Traynor AE. Hematopoietic stem cell transplantation: a new therapy for autoimmune disease. *Stem Cells* 1999;17(6):366.
269. Wolfson N, von Reper A, Berings M, et al. Autologous haematopoietic stem-cell transplantation in four patients with refractory juvenile chronic arthritis. *Lancet* 1999;353(9150):550.
270. Brodsky RA, Senesbrenner LL, Jones RJ. Complete remission in severe aplastic anemia after high-dose cyclophosphamide without bone marrow transplantation. *Blood* 1996;87(2):491.
271. Nossati EC, Brodsky RA, Jones RJ, Grever MR, Anhalt GJ. Immunoblastic high-dose cyclophosphamide without stem cell rescue in paraneoplastic pemphigus: report of a case and review of this new therapy for severe autoimmune disease. *J Am Acad Dermatol* 1999;40(5):790.

clinical trials demonstrated significant activity against several tumor types, particularly testicular tumors, the severe renal and gastrointestinal toxicity caused by the drug nearly led to its abandonment. Cvitkovic et al.^{3,4} showed that these effects could be ameliorated, in part, by aggressive prehydration, which rekindled interest in its clinical use. Currently, cisplatin is curative in testicular cancer and significantly prolongs survival in combination regimens for ovarian cancer. The drug also has therapeutic benefit in head and neck, bladder, and lung cancer.⁵

The unique activity and toxicity profile observed with cisplatin has fueled the development of platinum analogues that are less toxic and more effective against a variety of tumor types, including those that have developed resistance to cisplatin. Two other platinum drugs are widely used: *cis*-diamminocyclobutanedicarboxylato platinum (II) (carboplatin) and 1,2-diaminocyclohexanecarboxylato platinum (II) (oxaliplatin). Several new analogues with unique activities are currently in various stages of clinical development. Continued progress in the development of superior analogues requires a thorough understanding of the chemical, biological, pharmacokinetic, and pharmacodynamic properties of this important class of drugs. A review of these properties is the focus of this chapter.

PLATINUM CHEMISTRY

Platinum exists primarily in either a 2+ or 4+ oxidation state. These oxidation states dictate the stereochemistry of the carrier ligands and leaving groups surrounding the platinum atom. Platinum (II) compounds exhibit a square planar geometry, whereas platinum (IV) compounds exhibit an octahedral geometry. Interconversion of the two oxidation states may readily occur. However, the kinetics of this reaction depend on the nature of the bound ligands. The nature of the ligands also determines the stability of the complex and the rate of substitution. For platinum (II) compounds, the rate of substitution of a ligand is strongly influenced by the type of ligand located opposite to it. Therefore, ligands that are bound more strongly stabilize the moieties that are situated *trans* to it. For *cis*-diamminedichloroplatinum (II), the two chloride ligands are prone to substitution, whereas substitution of the amino groups is thermodynamically unfavorable.⁶ The stereochemistry of platinum complexes is critical to their antitumor activity, as ev-